

Lucia Vergauwen, Evelyn Stinckens, Daniel L. Villeneuve, Dries Knapen

Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation

https://dx.doi.org/10.1787/9f651c16-en



Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation

Series on Adverse Outcome Pathways No. 25

AOP No. 158 in the AOP-Wiki platform

Foreword

This Adverse Outcome Pathway (AOP) on Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation, has been developed under the auspices of the OECD AOP Development Programme, overseen by the Extended Advisory Group on Molecular Screening and Toxicogenomics (EAGMST), which is an advisory group under the Working Party of the National Coordinators for the Test Guidelines Programme (WNT) and the Working Party on Hazard Assessment (WPHA).

The AOP has been reviewed for compliance with the AOP development principles following the EAGMST coaching approach. The scientific review was subsequently conducted by the UK National Centre for the 3Rs, following the OECD AOP review principles outlined in the Guidance Document on the scientific review of AOPs. This AOP was endorsed by the WNT and the WPHA on 3 August 2022.

Through endorsement of this AOP, the WNT and the WPHA express confidence in the scientific review process that the AOP has undergone and accept the recommendation of the EAGMST that the AOP be disseminated publicly. Endorsement does not necessarily indicate that the AOP is now considered a tool for direct regulatory application.

The OECD's Chemicals and Biotechnology Committee agreed to declassification of this AOP on 4 November 2022.

This document is being published under the responsibility of the OECD's Chemicals and Biotechnology Committee.

The outcome of the compliance check and of the scientific review are publicly available respectively in the <u>AOP Wiki</u> and the <u>eAOP Portal of the AOP Knowledge Base</u> at the following links: <u>[internal review]</u> [scientific review report].

Authors:

Lucia Vergauwen [1], [lucia.vergauwen@uantwerpen.be] Evelyn Stinckens [1], [evelyn.stinckens@uantwerpen.be] Dan Villeneuve [2], [villeneuve.dan@epa.gov] Dries Knapen [1], [dries.knapen@uantwerpen.be] [1]

 Zebrafishlab, Veterinary Physiology and Biochemistry, Department of Veterinary Sciences, University of Antwerp, Universiteitsplein 1, 2610 Wilrijk, Belgium
 United States Environmental Protection Agency, Mid-Continent Ecology Division, 6201 Congdon Blvd, Duluth, MN, USA

Abstract

This AOP describes the sequence of events leading from deiodinase inhibition to increased mortality via reduced anterior swim bladder inflation. Thyroid hormones (THs) are critical during development and disruption of the TH system can interfere with normal development. Three types of iodothyronine deiodinases (DIO1-3) have been described in vertebrates that activate or inactivate THs and are therefore important mediators of TH action. While type II deiodinase (DIO2) has thyroxine (T4) as a preferred substrate and is mostly important for converting T4 to the more biologically active triiodothyronine (T3), type I deiodinase is capable of both converting T4 into T3 and converting rT3 to the inactive thyroid hormone 3,3' T2. Inhibition of DIO1 thus reduces T3 levels.

However, partly because rT3, rather than T4, is the preferred substrate for DIO1, DIO1 inhibition is probably less important in causing reduced T3 levels when compared to DIO2 inhibition. Swim bladder inflation is known to be under TH control (Brown et al.,1988; Liu and Chan, 2002). Many fish species have a swim bladder which is a gas-filled organ that typically consists of two chambers (Robertson et al., 2007). The posterior chamber inflates during early development in the embryonic phase, while the anterior chamber inflates during buoyancy, and the anterior chamber has an additional role in hearing (Robertson et al., 2017). This AOP describes how inhibition of DIO1 reduces levels of T3, thereby prohibiting proper inflation of the anterior chamber. Due to its role in regulating buoyancy, this results in reduced swimming performance. Since reduced swimming performance resuls in a decreased ability to forage and avoid predators, this reduces chances of survival. The final adverse outcome is a decrease of the population level, the more specific and informative adverse outcome at the organismal level, increased mortality, is used in the AOP title. Support for this AOP is mainly based on chemical exposures in zebrafish and fathead minnows (Cavallin et al., 2017; Stinckens et al., 2017; Stinckens et al., 2020).

This AOP is part of a larger AOP network describing how decreased synthesis and/or decreased biological activation of THs leads to incomplete or improper inflation of the swim bladder, leading to reduced swimming performance, increased mortality and decreased population trajectory (Knapen et al., 2018; Knapen et al., 2020; Villeneuve et al., 2018). Other than the difference in deiodinase (DIO) isoform, the current AOP is identical to the corresponding AOP leading from DIO2 inhibition to increased mortality via anterior swim bladder inflation (https://aopwiki.org/aops/156). The overall importance of DIO1 versus DIO2 in fish is not exactly clear. DIO1 inhibitors are often also inhibitors of DIO2 (Olker et al., 2019; Stinckens et al. 2018). In the ToxCast DIO1 inhibition single concentration assay, 219 out of 1820 chemicals were positive and 177 of these were also positive for DIO2 inhibition (viewed on 5/7/2022). This complicates the distinction between the relative contribution of DIO1 and DIO2 inhibition to reduced swim bladder inflation. The current state of the art suggests that DIO2 is more important than DIO1 in regulating swim bladder inflation. Six out of seven DIO1 inhibitors impaired posterior chamber inflation, but almost all of these compounds also inhibit DIO2 (Stinckens et al., 2018)). Tetrachlorobisphenol A (TCBPA), the only compound that inhibits DIO1 and not DIO2, had no effect on the posterior swim bladder. Exposure to strong DIO2 inhibitors on the other hand affected posterior chamber inflation and/or surface area in all cases. Therefore AOP 156 may be of higher biological relevance compared to the AOP that is described here. Starting from reduced serum T3 levels, this AOP is also identical to the AOP leading from thyroperoxidase inhibition leading to increased mortality via reduced anterior swim bladder inflation (https://aopwiki.org/aops/159).

Table of contents

| Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim b inflation | bladder 1 |
|--|--|
| Foreword | 2 |
| Background | 7 |
| Graphical Representation | 8 |
| Summary of the AOP Events Key Event Relationships | 9 9 9 |
| Overall Assessment of the AOP Domain of Applicability Essentiality of the Key Events Evidence Assessment Quantitative Understanding | 10 10 11 11 12 |
| Considerations for Potential Applications of the AOP | 13 |
| Appendix 1 - MIE, KEs and AO List of MIEs in this AOP Event: 1009: Inhibition, Deiodinase 1 Evidence for Perturbation by Stressor Domain of Applicability Key Event Description How it is Measured or Detected List of Key Events in the AOP Event: 1003: Decreased, Triiodothyronine (T3) Domain of Applicability Key Event Description How it is Measured or Detected Event: 1007: Reduced, Anterior swim bladder inflation Domain of Applicability Key Event Description How it is Measured or Detected Event: 1007: Reduced, Anterior swim bladder inflation Domain of Applicability Key Event Description How it is Measured or Detected Event: 1005: Reduced, Swimming performance | 15 15 15 16 17 18 21 21 23 25 29 29 30 31 33 |
| Domain of Applicability | 33 |

| Key Event Description | 34 |
|--|------------|
| How it is Measured or Detected | 34 |
| List of Adverse Outcomes in this AOP | 35 |
| Event: 351: Increased Mortality | 35 |
| Domain of Applicability | 36 |
| Key Event Description | 36 |
| How it is Measured or Detected | 36 |
| Regulatory Significance of the AO | 37 |
| Event: 360: Decrease, Population growth rate | 38 |
| Domain of Applicability | 40 |
| Key Event Description | 40 |
| How it is Measured or Detected | 40 |
| Regulatory Significance of the AO | 40 |
| Appendix 2 - List of Key Event Relationships in the AOP | 42 |
| List of Adjacent Key Event Relationships | 42 |
| Relationship: 1037: Inhibition, Deiodinase 1 leads to Decreased, Triiodothyronine (T3) | 42 |
| Evidence Supporting Applicability of this Relationship | 42 |
| Key Event Relationship Description | 43 |
| Evidence Supporting this KER | 43 |
| Quantitative Understanding of the Linkage | 44 |
| Relationship: 1035: Decreased, Triiodothyronine (T3) leads to Reduced, Anterior swim bladder | |
| inflation | 47 |
| Evidence Supporting Applicability of this Relationship | 47 |
| Key Event Relationship Description | 48 |
| Evidence Supporting this KER | 48 |
| Relationship: 1034: Reduced, Anterior swim bladder inflation leads to Reduced, Swimming | |
| performance | 51 |
| Evidence Supporting Applicability of this Relationship | 51 |
| Key Event Relationship Description | 52 |
| Evidence Supporting this KER | 52 |
| Relationship: 2212: Reduced, Swimming performance leads to Increased Mortality | 56 |
| Evidence Supporting Applicability of this Relationship | 56 |
| Key Event Relationship Description | 57 57 |
| Evidence Supporting this KER Quantitative Understanding of the Linkage | 57 58 |
| | 59 |
| Relationship: 2013: Increased Mortality leads to Decrease, Population growth rate | 59 59 |
| Evidence Supporting Applicability of this Relationship Key Event Relationship Description | - 59 60 |
| Evidence Supporting this KER | 60 |
| | 00 |
| Annex 1: Weight of evidence evaluation table | 63 |
| | |

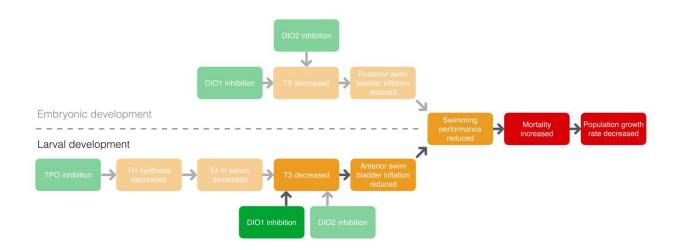
Background

The larger AOP network describing the effect of deiodinase and thyroperoxidase inhibition on swim bladder inflation consists of 5 AOPs:

- Deiodinase 2 inhibition leading to increased mortality via reduced posterior swim bladder inflation: <u>https://aopwiki.org/aops/155</u>
- Deiodinase 2 inhibition leading to increased mortality via reduced anterior swim bladder inflation: <u>https://aopwiki.org/aops/156</u>
- Deiodinase 1 inhibition leading to increased mortality via reduced posterior swim bladder inflation : https://aopwiki.org/aops/157
- Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation : https://aopwiki.org/aops/158
- Thyroperoxidase inhibition leading to increased mortality via reduced anterior swim bladder inflation: <u>https://aopwiki.org/aops/159</u>

The development of these AOPs was mainly based on a series of dedicated experiments (using a set of reference chemicals as prototypical stressors) in zebrafish and fathead minnow that form the core of the empirical evidence. Specific literature searches were used to add evidence from other studies, mainly in zebrafish and fathead minnow. No systematic review approach was applied.

Graphical Representation



Summary of the AOP

Events

Molecular Initiating Events (MIE), Key Events (KE), Adverse Outcomes (AO)

| Sequ ence | Ty pe | Event ID | Title | Short name |
|--------------|----------|-------------|--|--|
| 1 | MI E | 1009 | Inhibition, Deiodinase 1 | Inhibition, Deiodinase 1 |
| 2 | KE | 1003 | Decreased, Triiodothyronine (T3) | Decreased, Triiodothyronine (T3) |
| 3 | KE | 1007 | Reduced, Anterior swim bladder inflation | Reduced, Anterior swim bladder inflation |
| 4 | KE | 1005 | Reduced, Swimming performance | Reduced, Swimming performance |
| 5 | A O | 351 | Increased Mortality | Increased Mortality |
| 6 | A O | 360 | Decrease, Population growth rate | Decrease, Population growth rate |

Key Event Relationships

| Upstream Event | Relationship Type | Downstream Event | Evidence | Quantitative Understanding |
|--|----------------------|--|----------|-------------------------------|
| Inhibition, Deiodinase 1 | adjacent | Decreased, Triiodothyronine (T3) | Low | Low |
| Decreased, Triiodothyronine (T3) | adjacent | Reduced, Anterior swim bladder inflation | Moderate | Moderate |
| Reduced, Anterior swim bladder inflation | adjacent | Reduced, Swimming performance | Moderate | Low |
| Reduced, Swimming performance | adjacent | Increased Mortality | Moderate | Low |
| Increased Mortality | adjacent | Decrease, Population growth rate | Moderate | Moderate |

Overall Assessment of the AOP

The document in Annex 1 includes:

- Support for biological plausibility of KERs
- Support for essentiality of KEs
- Empirical support for KERs
- Dose and temporal concordance table covering the larger AOP network

Overall, the weight of evidence for the sequence of key events laid out in the AOP is moderate, and it should be noted that based on available evidence DIO2 seems to be more important than DIO1 in providing sufficient T3 for swim bladder inflation. The exact underlying mechanism of TH disruption leading to impaired swim bladder inflation is not exactly understood.

Domain of Applicability

Life Stage Applicability

| Life stage | Evidence |
|------------|----------|
| Larvae | High |

Taxonomic Applicability

| Term | Scientific Term | Evidence | Link |
|----------------|---------------------|----------|-------------|
| zebrafish | Danio rerio | High | <u>NCBI</u> |
| fathead minnow | Pimephales promelas | High | NCBI |

Sex Applicability

| Sex | Evidence |
|------------|----------|
| Unspecific | Moderate |

Taxonomic: Organogenesis of the swim bladder begins with an evagination from the gut. In physostomous fish, a connection between the swim bladder and the gut is retained. In physoclystous fish, once initial inflation by gulping atmospheric air at the water surface has occurred, the swim bladder is closed off from the digestive tract and swim bladder volume is regulated by gas secretion into the swim bladder (Woolley and Qin, 2010). This AOP is currently mainly based on experimental evidence from studies on zebrafish and fathead minnows, physostomous fish with a two-chambered swim bladder. This AOP is not applicable to fish that do not have a second swim bladder chamber that inflates during larval development, e.g., the Japanese rice fish or medaka (*Oryzias latipes*).

Life stage: The current AOP is only applicable to larval development, which is the period where the anterior swim bladder chamber inflates. In all life stages, the conversion of T4 into more biologically active T3 is

10 |

essential. Inhibition of DIO1therefore impacts swim bladder inflation in both early (<u>https://aopwiki.org/aops/157</u>) and late developmental life stages.

Sex: All key events in this AOP are plausibly applicable to both sexes. Sex differences are not often investigated in tests using early life stages of fish. For zebrafish and fathead minnow, it is currently unclear whether sex-related differences are important in determining the magnitude of the changes across the sequence of events in this AOP. Different fish species have different sex determination and differentiation strategies. Zebrafish do not have identifiable heteromorphic sex chromosomes and sex is determined by multiple genes and influenced by the environment (Nagabhushana and Mishra, 2016). Zebrafish are undifferentiated gonochorists since both sexes initially develop an immature ovary (Maack and Segner, 2003). Immature ovary development progresses until approximately the onset of the third week. Later, in female fish immature ovaries continue to develop further, while male fish undergo transformation of ovaries into testes. Final transformation into testes varies among male individuals, however finishes usually around 6 weeks post fertilization. Since the anterior chamber inflates around 21 days post fertilization in zebrafish, sex differences are expected to play a minor role in the current AOP. Fathead minnow gonad differentiation also occurs during larval development. Fathead minnows utilize a XY sex determination strategy and markers can be used to genotype sex in life stages where the sex is not yet clearly defined morphologically (Olmstead et al., 2011). Ovarian differentiation starts at 10 dph followed by rapid development (Van Aerle et al., 2004). At 25 dph germ cells of all stages up to the primary oocytes stage were present and at 120 dph, vitellogenic oocytes were present. The germ cells (spermatogonia) of the developing testes only entered meiosis around 90-120 dph. Mature testes with spermatozoa are present around 150 dph. Since the anterior chamber inflates around 14 days post fertilization (9 dph) in fathead minnows, sex differences are expected to play a minor role in the current AOP.

Essentiality of the Key Events

Overall, the support for essentiality of the KEs is low since there is limited direct evidence from specifically designed experimental studies illustrating essentiality. This includes evidence from combined DIO1 and DIO2 knockdown studies in zebrafish showing the link with reduced posterior chamber inflation, but anterior chamber inflation was not studied. There is additional indirect evidence that reduced thyroid hormone synthesis causes reduced anterior swim bladder inflation: Chopra et al. (2019) showed that knockdown of dual oxidase, important for thyroid hormone synthesis, reduced anterior swim bladder inflation. It should be noted that dual oxidase also plays a role in oxidative stress. There is no specific evidence for the essentiality of DIO1 inhibition independent of DIO2 inhibition and DIO2 seems more important than DIO1 in providing sufficient T3 for proper swim bladder inflation. Therefore the overall evidence for essentiality is considered low.

Evidence Assessment

Biological plausibility: see Table. Overall, the weight of evidence for the biological plausibility of the KERs in the AOP is moderate since there is empirical support for an association between the sets of KEs and the KERs are plausible based on analogy to accepted biological relationships, but scientific understanding is not completely established.

Empirical support: see Table. Overall, the empirical support for the KERs in the AOP is moderate since dependent changes in sets of KEs following exposure to several specific stressors has been demonstrated, with limited evidence for dose and temporal concordance and some uncertainties.

Quantitative Understanding

Quantitative understanding of this AOP is currently lacking.

Considerations for Potential Applications of the AOP

A growing number of environmental pollutants are known to adversely affect the thyroid hormone system, and major gaps have been identified in the tools available for the identification, and the hazard and risk assessment of these thyroid hormone disrupting chemicals. Villeneuve et al. (2014) discussed the relevance of swim bladder inflation as a potential key event and endpoint of interest in fish tests. Knapen et al. (2020) provide an example of how the adverse outcome pathway (AOP) framework and associated data generation can address current testing challenges in the context of fish early-life stage tests, and fish tests in general. While the AOP is only applicable to fish, some of the upstream KEs are relevant across vertebrates. The taxonomic domain of applicability call of the KEs can be found on the respective pages. A suite of assays covering all the essential biological processes involved in the underlying toxicological pathways can be implemented in a tiered screening and testing approach for thyroid hormone disruption in fish, using the levels of assessment of the OECD's Conceptual Framework for the Testing and Assessment of Endocrine Disrupting Chemicals as a guide. Specifically, for this AOP, deiodinase inhibition can be assessed using an in chemico assay, measurements of T3 levels could be added to the Fish Embryo Acute Toxicity (FET) test (OECD TG 236), the Fish Early Life Stage Toxicity (FELS) Test (OECD TG210) and the Fish Sexual Development Test (FSDT, OECD TG 234), and assessments of anterior chamber inflation and swimming performance could be added to the FELS Test and FSDT.

Thyroid hormone system disruption causes multiple unspecific effects. Addition of TH measurements could aid in increasing the diagnostic capacity of a battery of endpoints since they are specific to the TH system. A battery of endpoints would ideally include the MIE, the AO and TH levels as the causal link. It is also in this philosophy that TH measurements are currently being considered as one of the endpoints in project 2.64 of the OECD TG work plan, "Inclusion of thyroid endpoints in OECD fish Test Guidelines". While T3 measurements showed low levels of variation and were highly predictive of downstream effects in dedicated experiments to support this AOP, more variability may be present in other studies. Because of the rapid development in fish, it is important to compare T3 levels within specific developmental stages. For example, clear changes in T3 levels have been observed in zebrafish at 14, 21 and 32 dpf (Stinckens et al., 2020) and in fathead minnows at 4, 6, 10, 14, 18 and 21 dpf (Nelson et al., 2016; Cavallin et al., 2017) using liquid chromatography tandem mass spectrometry (LC–MS/MS).

The overall importance of DIO1 versus DIO2 in fish is not exactly clear. The current state of the art suggests that DIO2 is more important than DIO1 in regulating swim bladder inflation. Therefore AOP 156 may be more relevant for applications compared to the AOP that is described here.

References

Brown, C.L., Doroshov, S.I., Nunez, J.M., Hadley, C., Vaneenennaam, J., Nishioka, R.S., Bern, H.A., 1988. MATERNAL TRIIODOTHYRONINE INJECTIONS CAUSE INCREASES IN SWIMBLADDER INFLATION AND SURVIVAL RATES IN LARVAL STRIPED BASS, MORONE-SAXATILIS. Journal of Experimental Zoology 248, 168-176.

Cavallin, J.E., Ankley, G.T., Blackwell, B.R., Blanksma, C.A., Fay, K.A., Jensen, K.M., Kahl, M.D., Knapen, D., Kosian, P.A., Poole, S.T., Randolph, E.C., Schroeder, A.L., Vergauwen, L., Villeneuve, D.L., 2017. Impaired swim bladder inflation in early life stage fathead minnows exposed to a deiodinase inhibitor, iopanoic acid. Environmental Toxicology and Chemistry 36, 2942-2952.

Godfrey, A., Hooser, B., Abdelmoneim, A., Horzmann, K.A., Freemanc, J.L., Sepulveda, M.S., 2017. Thyroid disrupting effects of halogenated and next generation chemicals on the swim bladder development of zebrafish. Aquatic Toxicology 193, 228-235.

Knapen, D., Angrish, M.M., Fortin, M.C., Katsiadaki, I., Leonard, M., Margiotta-Casaluci, L., Munn, S., O'Brien, J.M., Pollesch, N., Smith, L.C., Zhang, X.W., Villeneuve, D.L., 2018. Adverse outcome pathway networks I: Development and applications. Environmental Toxicology and Chemistry 37, 1723-1733.

Knapen, D., Stinckens, E., Cavallin, J.E., Ankley, G.T., Holbech, H., Villeneuve, D.L., Vergauwen, L., 2020. Toward an AOP Network-Based Tiered Testing Strategy for the Assessment of Thyroid Hormone Disruption. Environmental Science & Technology 54, 8491-8499.

Liu, Y.W., Chan, W.K., 2002. Thyroid hormones are important for embryonic to larval transitory phase in zebrafish. Differentiation 70, 36-45.

Maack, G., Segner, H., 2003. Morphological development of the gonads in zebrafish. Journal of Fish Biology 62, 895-906.

Nagabhushana A, Mishra RK. 2016. Finding clues to the riddle of sex determination in zebrafish. Journal of Biosciences. 41(1):145-155.

Olmstead AW, Villeneuve DL, Ankley GT, Cavallin JE, Lindberg-Livingston A, Wehmas LC, Degitz SJ. 2011. A method for the determination of genetic sex in the fathead minnow, pimephales promelas, to support testing of endocrine-active chemicals. Environmental Science & Technology. 45(7):3090-3095.

Robertson, G.N., McGee, C.A.S., Dumbarton, T.C., Croll, R.P., Smith, F.M., 2007. Development of the swimbladder and its innervation in the zebrafish, Danio rerio. Journal of Morphology 268, 967-985.

Stinckens, E., Vergauwen, L., Blackwell, B.R., Anldey, G.T., Villeneuve, D.L., Knapen, D., 2020. Effect of Thyroperoxidase and Deiodinase Inhibition on Anterior Swim Bladder Inflation in the Zebrafish. Environmental Science & Technology 54, 6213-6223.

van Aerle R, Runnalls TJ, Tyler CR. 2004. Ontogeny of gonadal sex development relative to growth in fathead minnow. Journal of Fish Biology. 64(2):355-369.

Villeneuve, D., Angrish, M., Fortin, M., Katsiadaki, I., Leonard, M., Margiotta-Casaluci, L., Munn, S., O'Brien, J., Pollesch, N., Smith, L., Zhang, X., Knapen, D., 2018. Adverse Outcome Pathway Networks II: Network Analytics. Environ Toxicol Chem doi: 10.1002/etc.4124.

Villeneuve, D., Volz, D.C., Embry, M.R., Ankley, G.T., Belanger, S.E., Leonard, M., Schirmer, K., Tanguay, R., Truong, L., Wehmas, L., 2014. Investigating alternatives to the fish early-life stage test: a strategy for discovering and annotating adverse outcome pathways for early fish development. Environmental Toxicology and Chemistry 33, 158-169.

Woolley, L.D., Qin, J.G., 2010. Swimbladder inflation and its implication to the culture of marine finfish larvae. Reviews in Aquaculture 2, 181-190.

14 |

Appendix 1 - MIE, KEs and AO

List of MIEs in this AOP

Event: 1009: Inhibition, Deiodinase 1

Short Name: Inhibition, Deiodinase 1

Key Event Component

| Process | Object | Action |
|--------------------|---------------------------------|-----------|
| catalytic activity | type I iodothyronine deiodinase | decreased |

AOPs Including This Key Event

| AOP ID and Name | Event T | уре |
|---|--------------------|------------|
| Aop:157 - Deiodinase 1 inhibition leading to increased mortality via reduced posterior swim bladder inflation | Molecular Event | Initiating |
| Aop:158 - Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation | Molecular Event | Initiating |
| Aop:189 - Type I iodothyronine deiodinase (DIO1) inhibition leading to altered amphibian metamorphosis | Molecular Event | Initiating |

Stressors

| | Name |
|------------------|------|
| iopanoic acid | |
| Propylthiouracil | |

Biological Context

Molecular

Level of Biological Organization

Evidence for Perturbation by Stressor

Overview for Molecular Initiating Event

Propylthiouracil (PTU) is the prototypical DIO1 inhibitor in mammals, although teleostean and amphibian DIO1 enzymes are less sensitive to inhibition by PTU (Orozco et al., 2003; Kuiper et al., 2006). DIO1 inhibitors are often also inhibitors of DIO2 (Olker et al., 2019; Stinckens et al. 2018). In the ToxCast DIO1 inhibition single concentration assay, 219 out of 1820 chemicals were positive and 177 of these were also positive for DIO2 inhibition (viewed on 5/7/2022). Olker et al. (2019) identified 22 DIO1-specific inhibitors using a human recombinant DIO1 enzyme (e.g., genistein, 6-methyl-2-thiouracil, sulfasalazine). Another well-known inhibitor of DIO1 (and DIO2 and 3) is iopanoic acid (IOP). Renko et al. (2003, 2015) pointed out

that IOP is actually a substrate of DIO1 (and DIO2 and 3) which is in line with its action as a competitive inhibotor. In fact, many compounds inhibit all three DIO isoforms. Olker et al. (2019) identified 93 compounds that inhibit DIOs 1, 2 and 3.

Domain of Applicability

Taxonomic Applicability

| Term | Scientific Term | Evidence | Links |
|-----------------------|-----------------------|----------|-------------|
| rat | Rattus norvegicus | High | <u>NCBI</u> |
| mouse | Mus musculus | Moderate | <u>NCBI</u> |
| pigs | Sus scrofa | Moderate | NCBI |
| Ovis orientalis aries | Ovis aries | Moderate | NCBI |
| fathead minnow | Pimephales promelas | Moderate | NCBI |
| killifish | Fundulus heteroclitus | Moderate | NCBI |
| gilthead bream | Sparus aurata | Moderate | NCBI |
| African clawed frog | Xenopus laevis | Moderate | NCBI |
| human | Homo sapiens | High | NCBI |
| Oreochromis niloticus | Oreochromis niloticus | Moderate | NCBI |
| zebrafish | Danio rerio | Moderate | NCBI |

Life Stage Applicability

| Life Stage | Evidence |
|-----------------|----------|
| All life stages | Moderate |

Sex Applicability

| Sex | Evidence |
|------------|----------|
| Unspecific | Moderate |

Taxonomic: Deiodination by DIO enzymes is known to exist in a wide range of vertebrates and invertebrates. Therefore, this KE is plausibly applicable across vertebrates. Studies reporting DIO1 inhibition have used human liver (Kuiper et al., 2006), human recombinant DIO1 enzyme (Olker et al., 2019), rat (*Rattus norvegicus*) liver (Klaren et al., 2005; Freyberger and Ahr, 2006; Kuiper et al., 2006; Pavelka, 2010) and thyroid gland (Ferreira et al., 2002), mouse (*Mus musculus*) brain (hernandez et al., 2006), hog (*Sus scrofa domesticus*) liver (Stinckens et al., 2018), sheep (*Ovis orientalis aries*) fetal hepatic, renal and perirenal adipose tissue (Forhead et al., 2006), tadpole (*Xenopus laevis*) liver (Kuiper et al., 2006), fathead minnow (*Pimephales promelas*) whole fish (Noyes et al., 2011), Nile tilapia (*Oreochromis niloticus*) liver (Walpita et al., 2007), Gilthead Seabream (*Sparus aurata*) kidney (Klaren et al., 2005), and killifish (*Fundulus heteroclitus*) liver (Orozco et al., 2003) among others. The latter teleostean DIO1 enzymes as well as amphibian enzymes differ from other vertebrate DIO1 enzymes in their lower sensitivity to propylthiouracil (PTU), a typical DIO1 inhibitor in mammals.

In mammals, DIO2 is thought to control the intracellular concentration of T3, while DIO1 is thought to be more important in determining systemic T3 levels (Marsili et al., 2011), and the same appears to be true for birds. However, this hypothesis has been challenged. For example, Maia et al. (2005) determined that in a normal physiological situation in humans the contribution of DIO2 to plasma T3 levels is twice that of DIO1.

16 |

Only in a hyperthyroid state was the contribution of DIO1 higher than that of DIO2. A DIO1 knockout mouse showed normal T3 levels and a normal general phenotype and DIO1 was rather found to play a role in limiting the detrimental effects of conditions that alter normal thyroid function, including hyperthyroidism and iodine deficiency (Schneider et al., 2006). van der Spek et al. concluded that the primary role of DIO1 in vivo is to degrade inactivated TH (van der Spek et al., 2017).

By contrast, DIO1 function in teleostean and amphibian T3 plasma regulation is less clear (Finnson et al. 1999, Kuiper et al. 2006). The presence of DIO1 in the liver of teleosts has been a controversial issue, and both the high level of DIO2 activity and its expression in the liver of teleosts are unique among vertebrates (Orozco and Valverde, 2005). This could explain why DIO2 inhibition seems to be more important than DIO1 inhibition in determining the adverse outcome in zebrafish (Stinckens et al., 2018).

Life stage: Deiodinase activity is important for all vertebrate life stages. Already during early embryonic development, deiodinase activity is needed to regulate thyroid hormone concentrations and coordinate developmental processes. However, the role of DIO1 and DIO2 seems to be distinct. The fact that DIO1 knockdown during zebrafish development only causes developmental defects when combined with DIO2 knockdown (Walpita et al., 2010), suggests that DIO1 is only important in cases of increased TH need during specific stages of development, as supported by increased expression during such stages (Vergauwen et al., 2018), and in cases of thyroid hormone depletion in fish. There can also be differences in sensitivity between sexes. There is evidence for sex- and age-differences of Dio1 expression in mice (Schomburg et al., 2007).

Sex: This KE is plausibly applicable to both sexes. Deiodinases are important for TH homeostasis and identical in both sexes. There can however be differences in sensitivity between sexes. There is evidence for sex- and age-differences of Dio1 expression in mice (Schomburg et al., 2007).

Key Event Description

Disruption of the thyroid hormone system is increasingly being recognized as an important toxicity pathway, as it can cause many adverse outcomes. Thyroid hormones do not only play an important role in the adult individual, but they are also critical during embryonic development. Thyroid hormones (THs) play an important role in a wide range of biological processes in vertebrates including growth, development, reproduction, cardiac function, thermoregulation, response to injury, tissue repair and homeostasis. Numerous chemicals are known to disturb thyroid function, for example by inhibiting thyroperoxidase (TPO) or deiodinase (DIO), upregulating excretion pathways or modifying gene expression. The two major thyroid hormones are triiodothyronine (T3) and thyroxine (T4), both iodinated derivatives of tyrosine. Most TH actions depend on the binding of T3 to its nuclear receptors. Active and inactive THs are tightly regulated by enzymes called iodothyronine deiodinases (DIO). The activation occurs via outer ring deiodination (ORD), i.e. removing iodine from the outer, phenolic ring of T4 to form T3, while inactivation occurs via inner ring deiodination (IRD), i.e. removing iodine from the inner tyrosol ring of T4 or T3.

Three types of iodothyronine deiodinases (DIO1-3) have been described in vertebrates that activate or inactivate THs and are therefore important mediators of TH action. All deiodinases are integral membrane proteins of the thioredoxin superfamily that contain selenocysteine in their catalytic centre. Type I deiodinase is capable of both ORD and IRD, including the conversion of T4 into T3, as well as the conversion of reverse T3 (rT3) to 3,3'-Diiodothyronine (3,3' T2) rT3, rather than T4, is the preferred substrate for DIO1. furthermore, DIO1 has a very high Km (µM range, compared to nM range for DIO2) (Darras and Van Herck, 2012). Type II deiodinase (DIO2) is only capable of ORD activity with T4 as a preferred substrate (i.e., activation of T4 to T3). DIO3 can inner ring deiodinate T4 and T3 to the inactive forms of THs, rT3 and 3,3'-T2 respectively.

DIO1 is a plasma membrane protein with its catalytic domain facing the cytosol. The relative contribution of the DIOs to thyroid hormone levels varies amongst species, developmental stages and tissues.

How it is Measured or Detected

At this time, there are no approved OECD or EPA guideline protocols for measurement of DIO inhibition. Deiodination is the major pathway regulating T3 bioavailability in mammalian tissues. In vitro assays can be used to examine inhibition of deiodinase 1 (DIO1) activity upon exposure to thyroid disrupting compounds.

Several methods for deiodinase activity measurements are available. A first in vitro assay measures deiodinase activities by quantifying the radioactive iodine release from iodine-labelled substrates, depending on the preferred substrates of the isoforms of deiodinases (Ferreira et al., 2002; Forhead et al., 2006; Freyberger and Ahr, 2006; Pavelka, 2010; Stinckens et al., 2018). Another assay uses a chromatographybased method coupled to mass spectroscopy to measure products of thyroxin resulting from deiodinase type-1 activity (Butt et al., 2011). A colorimetric method (Renko et al., 2012), the Sandell-Kolthoff method, that measures the release of iodine from T4 is also available. Each of these assays requires a source of deiodinase which can be obtained for example using unexposed pig liver tissue (available from slaughterhouses) or rat liver tissue. Renko et al. (2015), Hornung et al. (2018) and Olker et al. (2019) on the other hand used an adenovirus expression system to produce the DIO1 enzyme and developed an assay for nonradioactive measurement of iodide released using the Sandell-Kolthoff method, a photometric method based on Ce4+ reduction (Renko et al., 2012), in a 96-well plate format. This assay was then used to screen the ToxCast Phase 1 chemical library. The specific synthesis of DIO1 through the adenovirus expression system provides an important advantage over other methods where activity of the different deiodinase isoforms needs to be distinguished in other ways, such as based on differences in enzyme kinetics.

Measurements of in vivo deiodinase activity in tissues collected from animal experiments are scarce. Noyes et al. (2011) showed decreased rate of outer ring deiodination (mediated by DIO1 and DIO2) in whole fish microsomes after exposure to BDE-209. After incubation with the substrate, thyroid hormone levels were measured using LC-MS/MS. Houbrechts et al. (2016) confirmed decreased DIO1 activity in a DIO1-DIO2 knockdown zebrafish at the ages of 3 and 7 days post fertilization. Decreased T3 levels are often used as evidence of DIO inhibition, for example after exposure to iopanoic acid, in fish species such as zebrafish (Stinckens et al., 2020) and fathead minnow (Cavallin et al., 2017). It should be noted that it is difficult to make the distinction between decreased T3 levels caused by outer ring deiodination mediated by DIO2 inhibition or DIO1 inhibition. Renko et al. (2022) showed tissue-specific changes in DIO1 activity in hyper-and hypothyroid mice.

References

Butt, C.M., Wang, D.L., Stapleton, H.M., 2011. Halogenated Phenolic Contaminants Inhibit the In Vitro Activity of the Thyroid-Regulating Deiodinases in Human Liver. Toxicological Sciences 124, 339-347.

Cavallin JE, Ankley GT, Blackwell BR, Blanksma CA, Fay KA, Jensen KM, Kahl MD, Knapen D, Kosian PA, Poole ST et al. 2017. Impaired swim bladder inflation in early life stage fathead minnows exposed to a deiodinase inhibitor, iopanoic acid. Environmental Toxicology and Chemistry. 36(11):2942-2952.

Darras, V.M., Van Herck, S.L.J., 2012. Iodothyronine deiodinase structure and function: from ascidians to humans. Journal of Endocrinology 215, 189-206.

Ferreira, A.C.F., Lisboa, P.C., Oliveira, K.J., Lima, L.P., Barros, I.A., Carvalho, D.P., 2002. Inhibition of thyroid type 1 deiodinase activity by flavonoids. Food and Chemical Toxicology 40, 913-917.

Finnson, K.W., McLeese, J.M., Eales, J.G., 1999. Deiodination and deconjugation of thyroid hormone conjugates and type I deiodination in liver of rainbow trout, Oncorhynchus mykiss. General and Comparative Endocrinology 115, 387-397.

Forhead, A.J., Curtis, K., Kaptein, E., Visser, T.J., Fowden, A.L., 2006. Developmental control of iodothyronine deiodinases by cortisol in the ovine fetus and placenta near term. Endocrinology 147, 5988-5994.

Freyberger, A., Ahr, H.J., 2006. Studies on the goitrogenic mechanism of action of N,N,N',N'- tetramethylthiourea. Toxicology 217, 169-175.

Hernandez, A., Martinez, M.E., Fiering, S., Galton, V.A., St Germain, D., 2006. Type 3 deiodinase is critical for the maturation and function of the thyroid axis. J Clin Invest 116, 476-484.

Hornung, M.W., Korte, J.J., Olker, J.H., Denny, J.S., Knutsen, C., Hartig, P.C., Cardon, M.C., Degitz, S.J., 2018. Screening the ToxCast Phase 1 Chemical Library for Inhibition of Deiodinase Type 1 Activity. Toxicological Sciences 162, 570-581.

Houbrechts AM, Vergauwen L, Bagci E, Van Houcke J, Heijlen M, Kulemeka B, Hyde DR, Knapen D, Darras VM. 2016. Deiodinase knockdown affects zebrafish eye development at the level of gene expression, morphology and function. Molecular and Cellular Endocrinology. 424(C):81-93.

Klaren, P.H.M., Haasdijk, R., Metz, J.R., Nitsch, L.M.C., Darras, V.M., Van der Geyten, S., Flik, G., 2005. Characterization of an iodothyronine 5 '-deiodinase in gilthead seabream (Sparus auratus) that is inhibited by dithiothreitol. Endocrinology 146, 5621-5630.

Kuiper, G., Klootwijk, W., Dubois, G.M., Destree, O., Darras, V.M., Van der Geyten, S., Demeneix, B., Visser, T.J., 2006. Characterization of recombinant Xenopus laevis type I iodothyronine deiodinase: substitution of a proline residue in the catalytic center by serine (Pro132Ser) restores sensitivity to 6-propyl-2-thiouracil. Endocrinology 147, 3519-3529.

Marsili, A., Zavacki, A.M., Harney, J.W., Larsen, P.R., 2011. Physiological role and regulation of iodothyronine deiodinases: A 2011 update. Journal of Endocrinological Investigation 34, 395-407.

Noyes, P.D., Hinton, D.E., Stapleton, H.M., 2011. Accumulation and Debromination of Decabromodiphenyl Ether (BDE-209) in Juvenile Fathead Minnows (Pimephales promelas) Induces Thyroid Disruption and Liver Alterations. Toxicological Sciences 122, 265-274.

Olker, J.H., Korte, J.J., Denny, J.S., Hartig, P.C., Cardon, M.C., Knutsen, C.N., Kent, P.M., Christensen, J.P., Degitz, S.J., Hornung, M.W., 2019. Screening the ToxCast Phase 1, Phase 2, and e1k Chemical Libraries for Inhibitors of Iodothyronine Deiodinases. Toxicological Sciences 168, 430-442.

Orozco, A., Valverde, R.C., 2005. Thyroid hormone deiodination in fish. Thyroid 15, 799-813.

Orozco, A., Villalobos, P., Jeziorski, M.C., Valverde, C., 2003. The liver of Fundulus heteroclitus expresses deiodinase type 1 mRNA. General and Comparative Endocrinology 130, 84-91.

Pavelka, S., 2010. Radiometric enzyme assays: development of methods for extremely sensitive determination of types 1, 2 and 3 iodothyronine deiodinase enzyme activities. Journal of Radioanalytical and Nuclear Chemistry 286, 861-865.

Renko, K., Hoefig, C.S., Hiller, F., Schomburg, L., Kohrle, J., 2012. Identification of Iopanoic Acid as Substrate of Type 1 Deiodinase by a Novel Nonradioactive Iodide-Release Assay. Endocrinology 153, 2506-2513.

Renko K, Schäche S, Hoefig CS, Welsink T, Schwiebert C, Braun D, Becker NP, Köhrle J, and Schomburg L (2015). An improved nonradioactive screening method identifies genistein and xanthohumol as potent inhibitors of iodothyronine deiodinases. Thyroid 25, 962–968. [PubMed: 25962824]

Renko, K., Kerp, H., Pape, J., Rijntjes, E., Burgdorf, T., Führer, D., Köhrle, J. 2022. Tentative Application of a Streamlined Protocol to Determine Organ-Specific Regulations of Deiodinase 1 and Dehalogenase Activities as Readouts of the Hypothalamus-Pituitary-Thyroid-Periphery-Axis. Frontiers in Toxicology 4, 10.3389/ftox.2022.822993

Schneider, M.J., Fiering, S.N., Thai, B., Wu, S.Y., St Germain, E., Parlow, A.F., St Germain, D.L., Galton, V.A., 2006. Targeted disruption of the type 1 selenodeiodinase gene (Dio1) results in marked changes in thyroid hormone economy in mice. Endocrinology 147, 580-589.

Schomburg, L., Riese, C., Renko, K., Schweizer, U., 2007. Effect of age on sexually dimorphic selenoprotein expression in mice. Biological Chemistry 388, 1035-1041.

Stinckens, E., Vergauwen, L., Ankley, G.T., Blust, R., Darras, V.M., Villeneuve, D.L., Witters, H., Volz, D.C., Knapen, D., 2018. An AOP-based alternative testing strategy to predict the impact of thyroid hormone disruption on swim bladder inflation in zebrafish. Aquatic Toxicology 200, 1-12.

Stinckens E, Vergauwen L, Blackwell BR, Anldey GT, Villeneuve DL, Knapen D. 2020. Effect of thyroperoxidase and deiodinase inhibition on anterior swim bladder inflation in the zebrafish. Environmental Science & Technology. 54(10):6213-6223.

van der Spek, A.H., Fliers, E., Boelen, A., 2017. The classic pathways of thyroid hormone metabolism. Molecular and Cellular Endocrinology 458, 29-38.

Vergauwen, L., Cavallin, J.E., Ankley, G.T., Bars, C., Gabriels, I.J., Michiels, E.D.G., Fitzpatrick, K.R., Periz-Stanacev, J., Randolph, E.C., Robinson, S.L., Saari, T.W., Schroeder, A.L., Stinckens, E., Swintek, J., Van Cruchten, S.J., Verbueken, E., Villeneuve, D.L., Knapen, D., 2018. Gene transcription ontogeny of hypothalamic-pituitary-thyroid axis development in early-life stage fathead minnow and zebrafish. General and Comparative Endocrinology 266, 87-100.

Visser, T.J., Van Overmeeren, E., Fekkes, D., Docter, R., Hennemann, G. 1979. Inhibition of iodothyronine 5'-deiodinase by thioureylenes: structure-activity relationship. FEBS Letters, 103, 2.

Walpita, C.N., Crawford, A.D., Darras, V.M., 2010. Combined antisense knockdown of type 1 and type 2 iodothyronine deiodinases disrupts embryonic development in zebrafish (Danio rerio). Gen Comp Endocrinol 166, 134-141.

Walpita, C.N., Grommen, S.V., Darras, V.M., Van der Geyten, S., 2007. The influence of stress on thyroid hormone production and peripheral deiodination in the Nile tilapia (Oreochromis niloticus). Gen Comp Endocrinol 150, 18-25.

List of Key Events in the AOP

Event: 1003: Decreased, Triiodothyronine (T3)

Short Name: Decreased, Triiodothyronine (T3)

Key Event Component

| Process | Object | Action |
|----------------------------------|--------------------------|-----------|
| decreased triiodothyronine level | 3,3',5'-triiodothyronine | decreased |

AOPs Including This Key Event

| AOP ID and Name | Event Type |
|---|---------------|
| Aop:155 - Deiodinase 2 inhibition leading to increased mortality via reduced posterior swim bladder inflation | Key Event |
| Aop:156 - Deiodinase 2 inhibition leading to increased mortality via reduced anterior swim bladder inflation | Key Event |
| Aop:157 - Deiodinase 1 inhibition leading to increased mortality via reduced posterior swim bladder inflation | Key Event |
| Aop:158 - Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation | Key Event |
| Aop:189 - Type I iodothyronine deiodinase (DIO1) inhibition leading to altered amphibian metamorphosis | Key Event |
| Aop:159 - Thyroperoxidase inhibition leading to increased mortality via reduced anterior swim bladder inflation | Key Event |
| Aop:363 - Thyroperoxidase inhibition leading to increased mortality via altered retinal layer structure | Key Event |
| Aop:364 - Thyroperoxidase inhibition leading to increased mortality via decreased eye size | Key Event |
| Aop:365 - Thyroperoxidase inhibition leading to increased mortality via altered photoreceptor patterning | Key Event |

Biological Context

| | Level of Biological Organization |
|--------|----------------------------------|
| Tissue | |

Domain of Applicability

Taxonomic Applicability

| Term | Scientific Term | Evidence | Links |
|---------------------|---------------------|----------|-------------|
| zebrafish | Danio rerio | High | <u>NCBI</u> |
| fathead minnow | Pimephales promelas | High | NCBI |
| African clawed frog | Xenopus laevis | High | <u>NCBI</u> |

Life Stage Applicability

| Life Stage | Evidence |
|-----------------|----------|
| All life stages | High |
| | |
| | |

Sex Applicability

| Sex | Evidence |
|------------|----------|
| Unspecific | Moderate |

Taxonomic: The overall evidence supporting taxonomic applicability is strong. With few exceptions vertebrate species have T3 and T4 that are mostly bound to transport proteins in blood as well as T3 and T4 in tissues. Therefore, the current key event is plausibly applicable to vertebrates in general. Clear species differences exist in transport proteins (Yamauchi and Isihara, 2009). Specifically, the majority of supporting data for TH decreases come from rat studies and have been measured mostly in serum. The predominant iodothyronine binding protein in rat serum is transthyretin (TTR). TTR demonstrates a reduced binding affinity for T4 when compared with thyroxine binding globulin (TBG), the predominant serum binding protein for T4 in humans. This difference in serum binding protein affinity for THs is thought to modulate serum half-life for T4; the half-life of T4 in rats is 12-24 hr, whereas the half-life in humans is 5-9 days (Capen, 1997). While these species differences impact hormone half-life, possibly regulatory feedback mechanisms, and quantitative dose- response relationships, measurement of decreased THs is still regarded as a measurable key event causatively linked to downstream adverse outcomes.

Several studies have reported evidence of T3 decreases after exposure to TPO inhibitors and deiodinase inhibitors in early life stages of zebrafish (Stinckens et al., 2016; Stinckens et al., 2020; Wang et al., 2020) and fathead minnow (Nelson et al., 2016; Cavallin et al., 2017). Such measurements in fish early life stages are usually based on whole animal samples and do not allow for distinguishing between systemic and tissue TH alterations.

THs are evolutionarily conserved molecules present in all vertebrate species (Hulbert, 2000; Yen, 2001). Moreover, their crucial role in amphibian and lamprey metamorphoses (Manzon and Youson, 1997; Yaoita and Brown, 1990) as well as fish development, embryo-to-larval transition and larval-to-juvenile transition (Thienpont et al., 2011; Liu and Chan, 2002) is well established. Their role as environmental messenger via exogenous routes in echinoderms confirms the hypothesis that these molecules are widely distributed among the living organisms (Heyland and Hodin, 2004). However, the role of TH in the different species may differ depending on the expression or function of specific proteins (e.g., receptors or enzymes) that are related to TH function, and therefore extrapolation between species should be done with caution.

Life stage: THs are essential in all life stages, but decreases of TH levels are not applicable to all developmental phases. The earliest life stages of teleost fish rely on maternally transferred THs to regulate certain developmental processes until embryonic TH synthesis is active (Power et al., 2001). As a result, T4 levels are not expected to decrease in response to exposure to inhibitors of TH synthesis during these earliest stages of development. However, T3 levels are expected to decrease upon exposure to deiodinase inhibitors in any life stage, since maternal T4 needs to be activated to T3 by deiodinases similar to embryonically synthesized T4.

Sex: The KE is plausibly applicable to both sexes. THs are essential in both sexes and the components of the HPT- axis are identical in both sexes. There can however be sex-dependent differences in the sensitivity to the disruption of TH levels and the magnitude of the response. In humans, females appear more

22 |

susceptible to hypothyroidism compared to males when exposed to certain halogenated chemicals (Hernandez-Mariano et al., 2017; Webster et al., 2014). In adult zebrafish, Liu et al. (2019) showed sexdependent changes in TH levels and mRNA expression of regulatory genes including corticotropin releasing hormone (crh), thyroid stimulating hormone (tsh) and deiodinase 2 after exposure to organophosphate flame retardants. The underlying mechanism of any sex-related differences remains unclear.

Key Event Description

There are two biologically active thyroid hormones (THs), triiodothyronine (T3) and thyroxine (T4), and a few less active iodothyronines (rT3, 3,5-T2), which are all derived from the modification of tyrosine molecules (Hulbert, 2000). However, the plasma concentrations of the other iodothyronines are significantly lower than those of T3 and T4. The different iodothyronines are formed by the sequential outer or inner ring monodeiodination of T4 and T3 by the deiodinating enzymes, Dio1, Dio2, and Dio3 (Gereben et al., 2008). Deiodinase structure is considered to be unique, as THs are the only molecules in the body that incorporate iodide.

The circulatory system serves as the major transport and delivery system for THs from synthesis in the gland to delivery to tissues. The majority of THs in the blood are bound to transport proteins (Bartalena and Robbins, 1993). In humans, the major transport proteins are TBG (thyroxine binding globulin), TTR (transthyretin) and albumin. The percent bound to these proteins in adult humans is about 75, 15 and 10 percent, respectively (Schussler 2000). Unbound (free) hormones are approximately 0.03 and 0.3 percent for T4 and T3, respectively. In serum, it is the free form of the hormone that is active.

There are major species differences in the predominant binding proteins and their affinities for THs (see section below on Taxonomic applicability). However, there is broad agreement that changes in serum concentrations of THs is diagnostic of thyroid disease or chemical-induced disruption of thyroid homeostasis (Zoeller et al., 2007).

It is notable that the changes measured in the free TH concentration reflect mainly the changes in the serum transport proteins rather than changes in the thyroid status. These thyroid-binding proteins serve as hormonal storage which ensures their even and constant distribution in the different tissues, while they protect the most sensitive ones in the case of severe changes in thyroid availability, like in thyroidectomies (Obregon et al., 1981). Initially, it was believed that all of the effects of TH were mediated by the binding of T3 to the thyroid nuclear receptors (TRa and TRb), a notion which is now questionable due to the increasing evidence that support the non-genomic action of TH (Davis et al., 2010, Moeller et al., 2006). Many non-nuclear TH binding sites have been identified to date and they usually lead to rapid cellular response in TH-effects (Bassett et al., 2003). Four types of thyroid hormone signaling have been defined (Anyetei-Anum et al., 2018): type 1 is the canonical pathway in which liganded TR binds directly to DNA; type 2 describes liganded TR tethered to chromatin-associated proteins, but not bound to DNA directly; type 3 suggests that liganded TR can exert its function without recruitment to chromatin in either the nucleus or cytoplasm; and type 4 proposes that thyroid hormone acts at the plasma membrane or in the cytoplasm without binding TR, a mechanism of action that is emerging as a key component of thyroid hormone signaling.

The production of THs in the thyroid gland and the circulation levels in the bloodstream are self-controlled by an efficiently regulated feedback mechanism across the Hypothalamus-Pituitary-Thyroid (HPT) axis. TH levels are regulated, not only in the plasma level, but also in the individual cell level, to maintain homeostasis. This is succeeded by the efficient regulatory mechanism of the thyroid hormone axis which consists of the following: (1) the hypothalamic secretion of the thyrotropin-releasing hormone (TRH), (2) the thyroid-stimulating hormone (TSH) secretion from the anterior pituitary, (3) hormonal transport by the plasma binding proteins, (4) cellular uptake mechanisms in the cell level, (5) intracellular control of TH concentration by the deiodinating mechanism (6) transcriptional function of the nuclear thyroid hormone receptor and (7) in the fetus, the transplacental passage of T4 and T3 (Cheng et al., 2010).

In regards to the brain, the TH concentration involves also an additional level of regulation, namely the hormonal transport through the Blood Brain Barrier (BBB) (Williams, 2008). The TRH and the TSH regulate the production of thyroid hormones. Less T3 (the biologically more active TH) than T4 is produced by the thyroid gland. The rest of the required amount of T3 is produced by outer ring deiodination of T4 by the deiodinating enzymes D1 and D2 (Bianco et al., 2006), a process which takes place mainly in liver and kidneys but also in other target organs such as in the brain, the anterior pituitary, brown adipose tissue, thyroid and skeletal muscle (Gereben et al., 2008; Larsen, 2009). Both hormones exert their action in almost all tissues of mammals and they are acting intracellularly, and thus the uptake of T3 and T4 by the target cells is a crucial step of the overall pathway. The trans-membrane transport of TH is performed mainly through transporters that differ depending on the cell type (Hennemann et al., 2001; Friesema et al., 2005; Visser et al., 2008). Many transporter proteins have been identified to date. The monocarboxylate transporters (Mct8, Mct10) and the anion-transporting polypeptide (OATP1c1) show the highest degree of affinity towards TH (Jansen et al., 2005) and mutations in these genes have pathophysiological effects in humans (Bernal et al., 2015). Unlike humans with an MCT8 deficiency, MCT8 knockout mice do not have neurological impairment. One explanation for this discrepancy could be differences in expression of the T4 transporter OATP1C1 in the blood-brain barrier. This shows that cross-species differences in the importance of specific transporters may occur.

T3 and T4 have significant effects on normal development, neural differentiation, growth rate and metabolism (Yen, 2001; Brent, 2012; Williams, 2008), with the most prominent ones to occur during the fetal development and early childhood. The clinical features of hypothyroidism and hyperthyroidism emphasize the pleiotropic effects of these hormones on many different pathways and target organs. The thyroidal actions though are not only restricted to mammals, as their high significance has been identified also for other vertebrates, with the most well-studied to be the amphibian metamorphosis (Furlow and Neff, 2006). The importance of the thyroid-regulated pathways becomes more apparent in iodine deficient areas of the world, where a higher rate of cretinism and growth retardation has been observed and linked to decreased TH levels (Gilbert et al., 2012). Another very common cause of severe hypothyroidism in human is the congenital hypothyroidism, but the manifestation of these effects is only detectable in the lack of adequate treatment and is mainly related to neurological impairment and growth retardation (Glinoer, 2001), emphasizing the role of TH in neurodevelopment in all above cases. In adults, the thyroid-related effects are mainly linked to metabolic activities, such as deficiencies in oxygen consumption, and in the metabolism of the vitamin, proteins, lipids and carbohydrates, but these defects are subtle and reversible (Oetting and Yen, 2007). Blood tests to detect the amount of thyroid hormone (T4) and thyroid stimulating hormone (TSH) are routinely done for newborn babies for the diagnosis of congenital hypothyroidism at the earliest stage possible.

Although the components of the thyroid hormone system as well as thyroid hormone synthesis and action are highly conserved across vertebrates, there are some taxon-specific considerations.

Although the HPT axis is highly conserved, there are some differences between fish and mammals (Blanton and Specker, 2007; Deal and Volkoff, 2020). For example, in fish, corticotropin releasing hormone (CRH) often plays a more important role in regulating thyrotropin (TSH) secretion by the pituitary and thus thyroid hormone synthesis compared to TSH-releasing hormone (TRH). TTRs from fish have low sequence identity with human TTR, for example seabream TTR has 54% sequence identity with human TTR but the only amino acid difference within the thyroxine-binding site is the conservative substitution of Ser117 in human TTR to Thr117 in seabream TTR (Santos and Power, 1999; Yamauchi et al., 1999; Eneqvist et al., 2004). In vitro binding experiments showed that TH disrupting chemicals bind with equal or weaker affinity to seabream TTR than to the human TTR with polar TH disrupting chemicals, in particular, showing a more than 500-fold lower affinity for seabream TTR compared to human TTR (Zhang et al., 2018).

Zebrafish and fathead minnows are oviparous fish species in which maternal thyroid hormones are transferred to the eggs and regulate early embryonic developmental processes during external (versus intrauterine in mammals) development (Power et al., 2001; Campinho et al., 2014; Ruuskanen and Hsu, 2018) until embryonic thyroid hormone synthesis is initiated. Maternal transfer of thyroid hormones, both T4 and T3, to the eggs has been demonstrated in zebrafish (Walpita et al., 2007; Chang et al., 2012) and fathead minnows (Crane et al., 2004; Nelson et al., 2016).

Several studies have reported evidence of T3 decreases after exposure to TPO inhibitors and deiodinase inhibitors in early life stages of zebrafish (Stinckens et al., 2016; Stinckens et al., 2020; Wang et al., 2020) and fathead minnow (Nelson et al., 2016; Cavallin et al., 2017).

How it is Measured or Detected

T3 and T4 can be measured as free (unbound) or total (bound + unbound) in serum, or in tissues. Free hormone are considered more direct indicators of T4 and T3 activities in the body. The majority of T3 and T4 measurements are made using either RIA or ELISA kits. In animal studies, total T3 and T4 are typically measured as the concentrations of free hormone are very low and difficult to detect.

Historically, the most widely used method in toxicology is RIA. The method is routinely used in rodent endocrine and toxicity studies. The ELISA method has become more routine in rodent studies. The ELISA method is a commonly used as a human clinical test method.

Recently, analytical determination of iodothyronines (T3, T4, rT3, T2) and their conjugates through methods employing HPLC and mass spectrometry have become more common (DeVito et al., 1999; Miller et al., 2009; Hornung et al., 2015; Nelson et al., 2016; Stinckens et al., 2016).

Any of these measurements should be evaluated for fit-for-purpose, relationship to the actual endpoint of interest, repeatability, and reproducibility. All three of the methods summarized above would be fit-for-purpose, depending on the number of samples to be evaluated and the associated costs of each method. Both RIA and ELISA measure THs by an indirect methodology, whereas analytical determination is the most direct measurement available. All of these methods, particularly RIA, are repeatable and reproducible.

In fish early life stages most evidence for the ontogeny of TH synthesis comes from measurements of wholebody TH levels and using LC-MS techniques (Hornung et al., 2015) are increasingly used to accurately quantify whole-body TH levels (Nelson et al., 2016; Stinckens et al., 2016; Stinckens et al., 2020).

References

Anyetei-Anum, C.S., Roggero, V.R., Allison, L.A., 2018. Thyroid hormone receptor localization in target tissues. Journal of Endocrinology 237, R19-R34.

Bartalena L, Robbins J.Thyroid hormone transport proteins.Clin Lab Med. 1993 Sep;13(3):583-98.

Bassett JH, Harvey CB, Williams GR. (2003). Mechanisms of thyroid hormone receptor-specific nuclear and extra nuclear actions. Mol Cell Endocrinol. 213:1-11.

Bernal, J., Guadano-Ferraz, A., Morte, B., 2015. Thyroid hormone transporters-functions and clinical implications. Nature Reviews Endocrinology 11, 406-417.

Bianco AC, Kim BW. (2006). Deiodinases: implications of the local control of thyroid hormone action. J Clin Invest. 116: 2571–2579.

Blanton ML, Specker JL. 2007. The hypothalamic-pituitary-thyroid (hpt) axis in fish and its role in fish development and reproduction. Crit Rev Toxicol. 37(1-2):97-115.

Brent GA. (2012). Mechanisms of thyroid hormone action. J Clin Invest. 122: 3035-3043.

Campinho MA, Saraiva J, Florindo C, Power DM. 2014. Maternal thyroid hormones are essential for neural development in zebrafish. Molecular Endocrinology. 28(7):1136-1149.

Cavallin JE, Ankley GT, Blackwell BR, Blanksma CA, Fay KA, Jensen KM, Kahl MD, Knapen D, Kosian PA, Poole ST et al. 2017. Impaired swim bladder inflation in early life stage fathead minnows exposed to a deiodinase inhibitor, iopanoic acid. Environmental Toxicology and Chemistry. 36(11):2942-2952.

Chang J, Wang M, Gui W, Zhao Y, Yu L, Zhu G. 2012. Changes in thyroid hormone levels during zebrafish development. Zoological Science. 29(3):181-184.

Cheng SY, Leonard JL, Davis PJ. (2010). Molecular aspects of thyroid hormone actions. Endocr Rev. 31:139–170.

Crane HM, Pickford DB, Hutchinson TH, Brown JA. 2004. Developmental changes of thyroid hormones in the fathead minnow, pimephales promelas. General and Comparative Endocrinology. 139(1):55-60.

Davis PJ, Zhou M, Davis FB, Lansing L, Mousa SA, Lin HY. (2010). Mini-review: Cell surface receptor for thyroid hormone and nongenomic regulation of ion fluxes in excitable cells. Physiol Behav. 99:237–239.

Deal CK, Volkoff H. 2020. The role of the thyroid axis in fish. Frontiers in Endocrinology. 11.

DeVito M, Biegel L, Brouwer A, Brown S, Brucker-Davis F, Cheek AO, Christensen R, Colborn T, Cooke P, Crissman J, Crofton K, Doerge D, Gray E, Hauser P, Hurley P, Kohn M, Lazar J, McMaster S, McClain M, McConnell E, *Meier C, Miller R, Tietge J, Tyl R. (1999). Screening methods for thyroid hormone disruptors. Environ Health Perspect. 107:407-415.

Eales JG. (1997). Iodine metabolism and thyroid related functions in organisms lacking thyroid follicles: Are thyroid hormones also vitamins? Proc Soc Exp Biol Med. 214:302-317.

Eneqvist T, Lundberg E, Karlsson A, Huang SH, Santos CRA, Power DM, Sauer-Eriksson AE. 2004. High resolution crystal structures of piscine transthyretin reveal different binding modes for triiodothyronine and thyroxine. Journal of Biological Chemistry. 279(25):26411-26416.

Friesema EC, Jansen J, Milici C, Visser TJ. (2005). Thyroid hormone transporters. Vitam Horm. 70: 137–167.

Furlow JD, Neff ES. (2006). A developmental switch induced by thyroid hormone: Xenopus laevis metamorphosis. Trends Endocrinol Metab. 17:40–47.

Gereben B, Zavacki AM, Ribich S, Kim BW, Huang SA, Simonides WS, Zeöld A, Bianco AC. (2008). Cellular and molecular basis of deiodinase-regulated thyroid hormone signalling. Endocr Rev. 29:898–938.

Gilbert ME, Rovet J, Chen Z, Koibuchi N. (2012).Developmental thyroid hormone disruption: prevalence, environmental contaminants and neurodevelopmental consequences. Neurotoxicology. 33: 842-852.

Glinoer D. (2001).Potential consequences of maternal hypothyroidism on the offspring: evidence and implications. Horm Res. 55:109-114.

Hennemann G, Docter R, Friesema EC, de Jong M, Krenning EP, Visser TJ. (2001). Plasma membrane transport of thyroid hormones and its role in thyroid hormone metabolism and bioavailability. Endocr Rev. 22:451-476.

Hernandez-Mariano JA, Torres-Sanchez L, Bassol-Mayagoitia S, Escamilla-Nunez M, Cebrian ME, Villeda-Gutierrez EA, Lopez- Rodriguez G, Felix-Arellano EE, Blanco-Munoz J. 2017. Effect of exposure to p,p 'dde during the first half of pregnancy in the maternal thyroid profile of female residents in a mexican floriculture area. Environmental Research. 156:597-604. Heyland A, Hodin J. (2004). Heterochronic developmental shift caused by thyroid hormone in larval sand dollars and its implications for phenotypic plasticity and the evolution of non-feeding development. Evolution. 58: 524-538.

Heyland A, Moroz LL. (2005). Cross-kingdom hormonal signaling: an insight from thyroid hormone functions in marine larvae. J Exp Biol. 208:4355-4361.

Hornung, M.W., Kosian, P.A., Haselman, J.T., Korte, J.J., Challis, K., Macherla, C., Nevalainen, E., Degitz, S.J., 2015. In Vitro, Ex Vivo, and In Vivo Determination of Thyroid Hormone Modulating Activity of Benzothiazoles. Toxicological Sciences 146, 254-264.

Hulbert A J. (2000). Thyroid hormones and their effects: A new perspective. Biol Rev. 75: 519-631.

Jansen J, Friesema EC, Milici C, Visser TJ. (2005). Thyroid hormone transporters in health and disease. Thyroid. 15: 757-768.

Larsen PR. (2009).Type 2 iodothyronine deiodinase in human skeletal muscle: new insights into its physiological role and regulation. J Clin Endocrinol Metab. 94:1893-1895.

Liu XS, Cai Y, Wang Y, Xu SH, Ji K, Choi K. 2019. Effects of tris(1,3-dichloro-2-propyl) phosphate (tdcpp) and triphenyl phosphate (tpp) on sex-dependent alterations of thyroid hormones in adult zebrafish. Ecotoxicology and Environmental Safety. 170:25-32.

Liu YW, Chan WK. 2002. Thyroid hormones are important for embryonic to larval transitory phase in zebrafish. Differentiation. 70(1):36-45.

Manzon RG, Youson JH. (1997). The effects of exogenous thyroxine (T4) or triiodothyronine (T3), in the presence and absence of potassium perchlorate, on the incidence of metamorphosis and on serum T4 and T3 concentrations in larval sea lampreys (Petromyzon marinus L.). Gen Comp Endocrinol. 106:211-220.

Miller MD, Crofton KM, Rice DC, Zoeller RT. (2009). Thyroid-disrupting chemicals: interpreting upstream biomarkers of adverse outcomes. Environ Health Perspect. 117:1033-1041.

Moeller LC, Dumitrescu AM, Seo H, Refetoff S. (2006). Thyroid hormone mediated changes in gene expression can be initiated by cytosolic action of the thyroid hormone receptor β through the phosphatidylinositol 3-kinase pathway. NRS. 4:1-4.

Nelson, K., Schroeder, A., Ankley, G., Blackwell, B., Blanksma, C., Degitz, S., Flynn, K., Jensen, K., Johnson, R., Kahl, M., Knapen, D., Kosian, P., Milsk, R., Randolph, E., Saari, T., Stinckens, E., Vergauwen, L., Villeneuve, D., 2016. Impaired anterior swim bladder inflation following exposure to the thyroid peroxidase inhibitor 2-mercaptobenzothiazole part I: Fathead minnow. Aquatic Toxicology 173, 192-203.

Obregon MJ, Mallol J, Escobar del Rey F, Morreale de Escobar G. (1981). Presence of I-thyroxine and 3,5,3triiodo-I- thyronine in tissues from thyroidectomised rats. Endocrinology 109:908-913.

Oetting A, Yen PM. (2007). New insights into thyroid hormone action. Best Pract Res Clin Endocrinol Metab. 21:193–208.

Power DM, Llewellyn L, Faustino M, Nowell MA, Bjornsson BT, Einarsdottir IE, Canario AV, Sweeney GE. 2001. Thyroid hormones in growth and development of fish. Comp Biochem Physiol C Toxicol Pharmacol. 130(4):447-459.

Ruuskanen S, Hsu BY. 2018. Maternal thyroid hormones: An unexplored mechanism underlying maternal effects in an ecological framework. Physiological and Biochemical Zoology. 91(3):904-916.

Santos CRA, Power DM. 1999. Identification of transthyretin in fish (sparus aurata): Cdna cloning and characterisation. Endocrinology. 140(5):2430-2433.

Schussler, G.C. (2000). The thyroxine-binding proteins. Thyroid 10:141–149.

Stinckens E, Vergauwen L, Blackwell BR, Anldey GT, Villeneuve DL, Knapen D. 2020. Effect of thyroperoxidase and deiodinase inhibition on anterior swim bladder inflation in the zebrafish. Environmental Science & Technology. 54(10):6213-6223.

Stinckens, E., Vergauwen, L., Schroeder, A., Maho, W., Blackwell, B., Witters, H., Blust, R., Ankley, G., Covaci, A., Villeneuve, D., Knapen, D., 2016. Impaired anterior swim bladder inflation following exposure to the thyroid peroxidase inhibitor 2- mercaptobenzothiazole part II: Zebrafish. Aquatic Toxicology 173, 204-217.

Thienpont B, Tingaud-Sequeira A, Prats E, Barata C, Babin PJ, Raldua D. 2011. Zebrafish eleutheroembryos provide a suitable vertebrate model for screening chemicals that impair thyroid hormone synthesis. Environmental Science & Technology. 45(17):7525-7532

Visser WE, Friesema EC, Jansen J, Visser TJ. (2008). Thyroid hormone transport in and out of cells. Trends Endocrinol Metab. 19:50-56.

Walpita CN, Van der Geyten S, Rurangwa E, Darras VM. 2007. The effect of 3,5,3'-triiodothyronine supplementation on zebrafish (danio rerio) embryonic development and expression of iodothyronine deiodinases and thyroid hormone receptors. Gen Comp Endocrinol. 152(2-3):206-214.

Wang JX, Shi GH, Yao JZ, Sheng N, Cui RN, Su ZB, Guo Y, Dai JY. 2020. Perfluoropolyether carboxylic acids (novel alternatives to pfoa) impair zebrafish posterior swim bladder development via thyroid hormone disruption. Environment International. 134.

Webster GM, Venners SA, Mattman A, Martin JW. 2014. Associations between perfluoroalkyl acids (pfass) and maternal thyroid hormones in early pregnancy: A population-based cohort study. Environmental Research. 133:338-347.

Williams GR. (2008). Neurodevelopmental and neurophysiological actions of thyroid hormone. J Neuroendocrinol. 20:784–794.

Yamauchi K, Nakajima J, Hayashi H, Hara A. 1999. Purification and characterization of thyroid-hormonebinding protein from masu salmon serum - a homolog of higher-vertebrate transthyretin. European Journal of Biochemistry. 265(3):944-949.

Yamauchi K1, Ishihara A. Evolutionary changes to transthyretin: developmentally regulated and tissuespecific gene expression.FEBS J. 2009 Oct;276(19):5357-66.

Yaoita Y, Brown DD. (1990). A correlation of thyroid hormone receptor gene expression with amphibian metamorphosis. Genes Dev. 4:1917-1924.

Yen PM. (2001). Physiological and molecular basis of thyroid hormone action. Physiol Rev. 81:1097-1142.

Zhang J, Grundstrom C, Brannstrom K, Iakovleva I, Lindberg M, Olofsson A, Andersson PL, Sauer-Eriksson AE. 2018. Interspecies variation between fish and human transthyretins in their binding of thyroid-disrupting chemicals. Environmental Science & Technology. 52(20):11865-11874.

Zoeller RT, Tan SW, Tyl RW. General background on the hypothalamic-pituitary-thyroid (HPT) axis. Crit Rev Toxicol. 2007 Jan- Feb;37(1-2):11-53.

Event: 1007: Reduced, Anterior swim bladder inflation

Short Name: Reduced, Anterior swim bladder inflation

Key Event Component

| Process | Object | Action |
|------------------------|-------------------------------|-----------|
| swim bladder inflation | anterior chamber swim bladder | decreased |

AOPs Including This Key Event

| AOP ID and Name | Event Type |
|---|---------------|
| Aop:156 - Deiodinase 2 inhibition leading to increased mortality via reduced anterior swim bladder inflation | Key Event |
| Aop:158 - Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation | Key Event |
| Aop:159 - Thyroperoxidase inhibition leading to increased mortality via reduced anterior swim bladder inflation | Key Event |

Biological Context

| Level of Biological Organization | | |
|----------------------------------|------------|--|
| Organ | | |
| Organ term | | |
| | Organ term | |
| swim bladder | | |

Domain of Applicability

Taxonomic Applicability

| Term | Scientific Term | Evidence | Links |
|----------------|---------------------|----------|-------------|
| zebrafish | Danio rerio | High | <u>NCBI</u> |
| fathead minnow | Pimephales promelas | High | <u>NCBI</u> |

Life Stage Applicability

| Life Stage | Evidence |
|------------|----------|
| Larvae | High |

Sex Applicability

| Sex | Evidence |
|------------|----------|
| Unspecific | Moderate |

Taxonomic: Teleost fish can be divided in two groups according to swim bladder morphology: physoclistous (e.g., yellow perch, sea bass, striped bass, medaka) and physostomous (e.g., zebrafish and fathead minnow). Physostomous fish retain a duct between the digestive tract and the swim bladder during adulthood

allowing them to gulp air at the surface to fill the swim bladder. In contrast, in physoclistous fish, once initial inflation by gulping atmospheric air at the water surface has occurred, the swim bladder is closed off from the digestive tract and swim bladder volume is regulated by gas secretion into the swim bladder (Woolley and Qin, 2010). The evidence for impaired inflation of the anterior chamber of the swim bladder currently comes from work on zebrafish and fathead minnow (Stinckens et al., 2016; Nelson et al., 2016; Cavallin et al., 2017; Godfrey et al., 2017; Stinckens et al., 2020). While zebrafish and fathead minnows are physostomous fish with a two-chambered swim bladder, the Japanese rice fish or medaka (*Oryzias latipes*) is a physoclistous fish with a single chambered swim bladder that inflates during early development. The key event 'reduced anterior chamber inflation' is not applicable to such fish species. Therefore, the current key event is plausibly applicable to physostomous fish in general.

Life stage: The anterior chamber inflates during a specific developmental time frame. In zebrafish, the anterior chamber inflates around 21 days post fertilization (dpf) which is during the larval stage. In the fathead minnow, the anterior chamber inflates around 14 dpf, also during the larval stage. Therefore this KE is only applicable to the larval life stage.

Sex: This KE plausibly applicable to both sexes. Sex differences are not often investigated in tests using early life stages of fish. For zebrafish and fathead minnow, it is currently unclear whether sex-related differences are important in determining the magnitude of the changes in this KE. Different fish species have different sex determination and differentiation strategies. Zebrafish do not have identifiable heteromorphic sex chromosomes and sex is determined by multiple genes and influenced by the environment (Nagabhushana and Mishra, 2016). Zebrafish are undifferentiated gonochorists since both sexes initially develop an immature ovary (Maack and Segner, 2003). Immature ovary development progresses until approximately the onset of the third week. Later, in female fish immature ovaries continue to develop further, while male fish undergo transformation of ovaries into testes. Final transformation into testes varies among male individuals, however finishes usually around 6 weeks post fertilization. Since the anterior chamber inflates around 21 days post fertilization in zebrafish, sex differences are expected to play a minor role. Fathead minnow gonad differentiation also occurs during larval development. Fathead minnows utilize a XY sex determination strategy and markers can be used to genotype sex in life stages where the sex is not yet clearly defined morphologically (Olmstead et al., 2011). Ovarian differentiation starts at 10 dph followed by rapid development (Van Aerle et al., 2004). At 25 dph germ cells of all stages up to the primary oocytes stage were present and at 120 dph, vitellogenic oocytes were present. The germ cells (spermatogonia) of the developing testes only entered meiosis around 90-120 dph. Mature testes with spermatozoa are present around 150 dph. Since the anterior chamber inflates around 14 days post fertilization (9 dph) in fathead minnows, sex differences are expected to play a minor role in the current KE.

Key Event Description

The swim bladder of bony fish is evolutionary homologous to the lung (Zheng et al., 2011). The teleost swim bladder is a gas-filled structure that consists of two chambers, the posterior and anterior chamber. In zebrafish, the posterior chamber inflates around 96 h post fertilization (hpf) which is 2 days post hatch, and the anterior chamber inflates around 21 dpf. In fathead minnow, the posterior and anterior chamber inflate around 6 and 14 dpf respectively. Inflation of the anterior swim bladder chamber is part of the larval-to-juvenile transition in fish, together with the development of adult fins and fin rays, ossification of the axial skeleton, formation of an adult pigmentation pattern, scale formation, maturation and remodeling of organs including the lateral line, nervous system, gut and kidneys (McMenamin and Parichy, 2013).

The anterior chamber is formed by evagination from the cranial end of the posterior chamber (Robertson et al., 2007). Dumbarton et al. (2010) showed that the anterior chamber of zebrafish has particularly closely packed and highly organized bundles of muscle fibres, suggesting that contraction of these muscles would reduce swim bladder volume. While it had previously been suggested that the posterior chamber had a more important role as a hydrostatic organ, this implies high importance of the anterior chamber for buoyancy. The anterior chamber has an additional role in hearing (Bang et al., 2002). Weberian ossicles (the Weberian apparatus) connect the anterior chamber to the inner ear resulting in an amplification of sound waves. Reduced inflation of the anterior chamber may manifest itself as either a complete failure to inflate the chamber or reduced size of the chamber. Reduced size is often associated with a deviating morphology.

How it is Measured or Detected

In several fish species, inflation of the anterior chamber can be observed using a stereomicroscope because the larvae are still transparent during the larval stage. This is for example true for zebrafish and fathead minnow. Anterior chamber size can then be measured based on photographs with a calibrator.

References

Bang, P.I., Yelick, P.C., Malicko, J.J., Sewell, W.F. 2002. High-throughput behavioral screening method for detecting auditory response defects in zebrafish. Journal of Neuroscience Methods. 118, 177-187.

Cavallin, J.E., Ankley, G.T., Blackwell, B.R., Blanksma, C.A., Fay, K.A., Jensen, K.M., Kahl, M.D., Knapen, D., Kosian, P.A., Poole, S.T., Randolph, E.C., Schroeder, A.L., Vergauwen, L., Villeneuve, D.L., 2017. Impaired swim bladder inflation in early life stage fathead minnows exposed to a deiodinase inhibitor, iopanoic acid. Environmental Toxicology and Chemistry 36, 2942-2952.

Dumbarton, T.C., Stoyek, M., Croll, R.P., Smith, F.M., 2010. Adrenergic control of swimbladder deflation in the zebrafish (Danio rerio). J. Exp. Biol. 213,2536–2546, http://dx.doi.org/10.1242/jeb.039792.

Godfrey, A., Hooser, B., Abdelmoneim, A., Horzmann, K.A., Freemanc, J.L., Sepulveda, M.S., 2017. Thyroid disrupting effects of halogenated and next generation chemicals on the swim bladder development of zebrafish. Aquatic Toxicology 193, 228-235.

McMenamin, S.K., Parichy, D.M., 2013. Metamorphosis in Teleosts. Animal Metamorphosis 103, 127-165.

Nagabhushana A, Mishra RK. 2016. Finding clues to the riddle of sex determination in zebrafish. Journal of Biosciences. 41(1):145-155.

Nelson KR, Schroeder AL, Ankley GT, Blackwell BR, Blanksma C, Degitz SJ, Flynn KM, Jensen KM, Johnson RD, Kahl MD, Knapen D, Kosian PA, Milsk RY, Randolph EC, Saari T, Stinckens E, Vergauwen L, Villeneuve DL. 2016. Impaired anterior swim bladder inflation following exposure to the thyroid peroxidase inhibitor 2-mercaptobenzothiazole – Part I: fathead minnow. Aquatic Toxicology 173: 192-203.

Olmstead AW, Villeneuve DL, Ankley GT, Cavallin JE, Lindberg-Livingston A, Wehmas LC, Degitz SJ. 2011. A method for the determination of genetic sex in the fathead minnow, pimephales promelas, to support testing of endocrine-active chemicals. Environmental Science & Technology. 45(7):3090-3095.

Roberston, G.N., McGee, C.A.S., Dumbarton, T.C., Croll, R.P., Smith, F.M., 2007. Development of the swim bladder and its innervation in the zebrafish, Danio rerio. J. Morphol. 268, 967–985, http://dx.doi.org/10.1002/jmor.

Stinckens, E., Vergauwen, L., Blackwell, B.R., Anldey, G.T., Villeneuve, D.L., Knapen, D., 2020. Effect of Thyroperoxidase and Deiodinase Inhibition on Anterior Swim Bladder Inflation in the Zebrafish. Environmental Science & Technology 54, 6213-6223.

Stinckens, E., Vergauwen, L., Schroeder, A., Maho, W., Blackwell, B., Witters, H., Blust, R., Ankley, G., Covaci, A., Villeneuve, D., Knapen, D., 2016. Impaired anterior swim bladder inflation following exposure to the thyroid peroxidase inhibitor 2- mercaptobenzothiazole part II: Zebrafish. Aquatic Toxicology 173, 204-217.

van Aerle R, Runnalls TJ, Tyler CR. 2004. Ontogeny of gonadal sex development relative to growth in fathead minnow. Journal of Fish Biology. 64(2):355-369.

Woolley, L.D., Qin, J.G., 2010. Swimbladder inflation and its implication to the culture of marine finfish larvae. Reviews in Aquaculture 2, 181-190.

Zheng, W., Wang, Z., Collins, J.E., Andrews, R.M., Stemple, D., Gong, Z. 2011. Comparative transcriptome analyses indicate molecular homology of zebrafish swim bladder and mammalian lung. PLoS One 6, http://dx.doi.org/10.1371/

Event: 1005: Reduced, Swimming performance

Short Name: Reduced, Swimming performance

Key Event Component

| Process | Object | Action |
|--------------------|--------|-----------|
| aquatic locomotion | | decreased |

AOPs Including This Key Event

| AOP ID and Name | Event Type |
|---|---------------|
| Aop:155 - Deiodinase 2 inhibition leading to increased mortality via reduced posterior swim bladder inflation | Key Event |
| Aop:156 - Deiodinase 2 inhibition leading to increased mortality via reduced anterior swim bladder inflation | Key Event |
| Aop:157 - Deiodinase 1 inhibition leading to increased mortality via reduced posterior swim bladder inflation | Key Event |
| Aop:158 - Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation | Key Event |
| Aop:159 - Thyroperoxidase inhibition leading to increased mortality via reduced anterior swim bladder inflation | Key Event |
| Aop:242 - Inhibition of lysyl oxidase leading to enhanced chronic fish toxicity | Key Event |
| Aop:334 - Glucocorticoid Receptor Agonism Leading to Impaired Fin Regeneration | Key Event |

Biological Context

| Level of Biological Organization | |
|----------------------------------|--|
| Individual | |

Domain of Applicability

Taxonomic Applicability

| Term | Scientific Term | Evidence | Links |
|----------------|---------------------|----------|-------|
| zebrafish | Danio rerio | High | NCBI |
| teleost fish | teleost fish | High | NCBI |
| fathead minnow | Pimephales promelas | High | NCBI |

Life Stage Applicability

| Life Stage | Evidence |
|------------|----------|
| Larvae | Moderate |
| Juvenile | Moderate |
| Adult | Moderate |

Sex Applicability

| Sex | Evidence |
|------------|----------|
| Unspecific | Moderate |

Taxonomic: Importance of swimming performance for natural behaviour is generally applicable to fish and tho other taxa that rely on swimming to support vital behaviours.

Life stage: Importance of swimming performance for natural behaviour is generally applicable across all free-swimming life stages, i.e., post-embryonic life stages.

Sex: Importance of swimming performance for natural behaviour is generally applicable across sexes.

Key Event Description

Adequate swimming performance in fish is essential for behaviour such as foraging, predator avoidance and reproduction.

How it is Measured or Detected

For fish larvae, automated observation and tracking systems are commercially available and increasingly used for measuring swimming performance including distance travelled, duration of movements, swimming speed, etc. This kind of measurements is often included in publications describing effects of chemicals in zebrafish larvae (Hagenaars et al., 2014; Stinckens et al., 2016; Vergauwen et al., 2015).

For juvenile and adult fish, measurements of swim performance vary. However, in some circumstances, swim tunnels have been used to measure various data (Fu et al., 2013).

Little and Finger (1990) discussed swimming behavior as an indicator of sublethal toxicity in fish.

References

Fu C, Cao ZD, Fu SJ. 2013. The effects of caudal fin loss and regeneration on the swimming performance of three cyprinid fish species with different swimming capacitities. The Journal of Experimental Biology 216:3164-3174. doi:10.1242/jeb.084244

Hagenaars, A., Stinckens, E., Vergauwen, L., Bervoets, L., Knapen, D., 2014. PFOS affects posterior swim bladder chamber inflation and swimming performanceof zebrafish larvae. Aquat. Toxicol. 157, 225–235.

Little EE, Finger SE. 1990. Swimming behavior as an indicator of sublethal toxicity in fish. Environmental Toxicology and Chemistry. 9(1):13-19.

Stinckens, E., Vergauwen, L., Schroeder, A.L., Maho, W., Blackwell, B., Witter, H.,Blust, R., Ankley, G.T., Covaci, A., Villenueve, D.L., Knapen, D., 2016. Disruption of thyroid hormone balance after 2-mercaptobenzothiazole exposure causes swim bladder inflation impairment—part II: zebrafish. Aquat. Toxicol. 173:204-17.

Vergauwen, Lucia; Nørgaard Schmidt, Stine; Maho, Walid; Stickens, Evelyn; Hagenaars, An; Blust, Ronny; Mayer, Philipp; Covaci, Adrian; Knapen, Dries. 2014. A high throughput passive dosing format for the Fish Embryo Acute Toxicity test. Chemosphere. 139: 9-17.

34 |

List of Adverse Outcomes in this AOP

Event: 351: Increased Mortality

Short Name: Increased Mortality

Key Event Component

| Process | Object | Action |
|-----------|-----------|--------|
| mortality | increased | |

AOPs Including This Key Event

| AOP ID and Name | Event Type |
|---|--------------------|
| Aop:16 - Acetylcholinesterase inhibition leading to acute mortality | Adverse |
| Acres 0. Averal acrives abore al modulation loading to coute mortality | Outcome |
| Aop:96 - Axonal sodium channel modulation leading to acute mortality | Adverse Outcome |
| Aop:104 - Altered ion channel activity leading impaired heart function | Adverse |
| | Outcome |
| Aop:113 - Glutamate-gated chloride channel activation leading to acute mortality | Adverse Outcome |
| <u>Aop:160 - Ionotropic gamma-aminobutyric acid receptor activation mediated</u> neurotransmission inhibition leading to mortality | Adverse Outcome |
| Aop:161 - Glutamate-gated chloride channel activation leading to | Adverse |
| neurotransmission inhibition associated mortality | Outcome |
| Aop:138 - Organic anion transporter (OAT1) inhibition leading to renal failure and mortality | Adverse Outcome |
| Aop:177 - Cyclooxygenase 1 (COX1) inhibition leading to renal failure and | Adverse |
| mortality | Outcome |
| Aop:186 - unknown MIE leading to renal failure and mortality | Adverse Outcome |
| Aop:312 - Acetylcholinesterase Inhibition leading to Acute Mortality via Impaired Coordination & Movement | Adverse Outcome |
| Aop:320 - Binding of viral S-glycoprotein to ACE2 receptor leading to acute | Adverse |
| respiratory distress associated mortality | Outcome |
| Aop:155 - Deiodinase 2 inhibition leading to increased mortality via reduced | Adverse |
| posterior swim bladder inflation | Outcome |
| Aop:156 - Deiodinase 2 inhibition leading to increased mortality via reduced anterior swim bladder inflation | Adverse Outcome |
| Aop:157 - Deiodinase 1 inhibition leading to increased mortality via reduced | Adverse |
| posterior swim bladder inflation | Outcome |
| Aop:158 - Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation | Adverse Outcome |
| Aop:159 - Thyroperoxidase inhibition leading to increased mortality via reduced | Adverse |
| anterior swim bladder inflation | Outcome |
| Aop:363 - Thyroperoxidase inhibition leading to increased mortality via altered retinal layer structure | Adverse Outcome |
| Aop:377 - Dysregulated prolonged Toll Like Receptor 9 (TLR9) activation leading | Adverse |

| AOP ID and Name | Event Type |
|--|------------|
| to Acute Respiratory Distress Syndrome (ARDS) and Multiple Organ Dysfunction | Outcome |
| <u>(MOD)</u> | |
| Aop:364 - Thyroperoxidase inhibition leading to increased mortality via | Adverse |
| decreased eye size | Outcome |
| Aop:365 - Thyroperoxidase inhibition leading to increased mortality via altered | Adverse |
| photoreceptor patterning | Outcome |
| Aop:399 - Inhibition of Fyna leading to increased mortality via decreased eye size | Adverse |
| (Microphthalmos) | Outcome |
| Aop:413 - Oxidation and antagonism of reduced glutathione leading to mortality | Adverse |
| via acute renal failure | Outcome |
| Aop:410 - Repression of Gbx2 expression leads to defects in developing inner | Key Event |
| ear and consequently to increased mortality | |

Biological Context

| | Level of Biological Organization | |
|------------|----------------------------------|--|
| Population | | |

Domain of Applicability

Taxonomic Applicability

| Term | Scientific Term | Evidence | Links |
|-------------|-----------------|----------|-------------|
| all species | all species | High | <u>NCBI</u> |

Life Stage Applicability

| Life Stage | Evidence |
|-----------------|----------|
| All life stages | High |

Sex Applicability

| Sex | Evidence |
|------------|----------|
| Unspecific | Moderate |

All living things are susceptible to mortality.

Key Event Description

Increased mortality refers to an increase in the number of individuals dying in an experimental replicate group or in a population over a specific period of time.

How it is Measured or Detected

Mortality of animals is generally observed as cessation of the heart beat, breathing (gill or lung movement) and locomotory movements.

Mortality is typically measured by observation. Depending on the size of the organism, instruments such as microscopes may be used. The reported metric is mostly the mortality rate: the number of deaths in a given area or period, or from a particular cause.

Depending on the species and the study setup, mortality can be measured:

- in the lab by recording mortality during exposure experiments
- in dedicated setups simulating a realistic situation such as mesocosms or drainable ponds for aquatic species
- in the field, for example by determining age structure after one capture, or by capture-mark-recapture efforts. The latter is a method
- commonly used in ecology to estimate an animal population's size where it is impractical to count every individual.

Regulatory Significance of the AO

Increased mortality is one of the most common regulatory assessment endpoints, along with reduced growth and reduced reproduction.

Event: 360: Decrease, Population growth rate

Short Name: Decrease, Population growth rate

Key Event Component

| Process | Object | Action |
|------------------------|-------------------------|-----------|
| population growth rate | population of organisms | decreased |

AOPs Including This Key Event

| AOP ID and Name | Event Type |
|--|-----------------|
| Aop:23 - Androgen receptor agonism leading to reproductive dysfunction (in repeat-spawning fish) | Adverse Outcome |
| Aop:25 - Aromatase inhibition leading to reproductive dysfunction | Adverse Outcome |
| Aop:29 - Estrogen receptor agonism leading to reproductive dysfunction | Adverse Outcome |
| Aop:30 - Estrogen receptor antagonism leading to reproductive dysfunction | Adverse Outcome |
| Aop:100 - Cyclooxygenase inhibition leading to reproductive dysfunction via inhibition of female spawning behavior | Adverse Outcome |
| Aop:122 - Prolyl hydroxylase inhibition leading to reproductive dysfunction via increased HIF1 heterodimer formation | Adverse Outcome |
| Aop:123 - Unknown MIE leading to reproductive dysfunction via increased HIF- 1alpha transcription | Adverse Outcome |
| Aop:155 - Deiodinase 2 inhibition leading to increased mortality via reduced posterior swim bladder inflation | Adverse Outcome |
| Aop:156 - Deiodinase 2 inhibition leading to increased mortality via reduced anterior swim bladder inflation | Adverse Outcome |
| Aop:157 - Deiodinase 1 inhibition leading to increased mortality via reduced posterior swim bladder inflation | Adverse Outcome |
| Aop:158 - Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation | Adverse Outcome |
| Aop:159 - Thyroperoxidase inhibition leading to increased mortality via reduced anterior swim bladder inflation | Adverse Outcome |
| Aop:101 - Cyclooxygenase inhibition leading to reproductive dysfunction via inhibition of pheromone release | Adverse Outcome |
| Aop:102 - Cyclooxygenase inhibition leading to reproductive dysfunction via interference with meiotic prophase I /metaphase I transition | Adverse Outcome |
| Aop:63 - Cyclooxygenase inhibition leading to reproductive dysfunction | Adverse Outcome |
| Aop:103 - Cyclooxygenase inhibition leading to reproductive dysfunction via interference with spindle assembly checkpoint | Adverse Outcome |
| Aop:292 - Inhibition of tyrosinase leads to decreased population in fish | Adverse Outcome |
| Aop:310 - Embryonic Activation of the AHR leading to Reproductive failure, via epigenetic down-regulation of GnRHR | Adverse Outcome |
| Aop:16 - Acetylcholinesterase inhibition leading to acute mortality | Adverse Outcome |
| Aop:312 - Acetylcholinesterase Inhibition leading to Acute Mortality via Impaired Coordination & Movement | Adverse Outcome |
| Aop:334 - Glucocorticoid Receptor Agonism Leading to Impaired Fin Regeneration | Adverse Outcome |
| Aop:336 - DNA methyltransferase inhibition leading to population decline (1) | Adverse Outcome |
| Aop:337 - DNA methyltransferase inhibition leading to population decline (2) | Adverse Outcome |

| AOP ID and Name | Event Type |
|--|-----------------|
| Aop:338 - DNA methyltransferase inhibition leading to population decline (3) | Adverse Outcome |
| Aop:339 - DNA methyltransferase inhibition leading to population decline (4) | Adverse Outcome |
| Aop:340 - DNA methyltransferase inhibition leading to transgenerational effects (1) | Adverse Outcome |
| Aop:341 - DNA methyltransferase inhibition leading to transgenerational effects (2) | Adverse Outcome |
| Aop:289 - Inhibition of 5a-reductase leading to impaired fecundity in female fish | Adverse Outcome |
| Aop:297 - Inhibition of retinaldehyde dehydrogenase leads to population decline | Adverse Outcome |
| Aop:346 - Aromatase inhibition leads to male-biased sex ratio via impacts on gonad differentiation | Adverse Outcome |
| Aop:299 - Excessive reactive oxygen species production leading to population decline via reduced fatty acid beta-oxidation | Adverse Outcome |
| Aop:311 - Excessive reactive oxygen species production leading to population decline via mitochondrial dysfunction | Adverse Outcome |
| Aop:216 - Excessive reactive oxygen species production leading to population decline via follicular atresia | Adverse Outcome |
| Aop:238 - Excessive reactive oxygen species production leading to population decline via lipid peroxidation | Adverse Outcome |
| Aop:326 - Thermal stress leading to population decline (3) | Adverse Outcome |
| Aop:325 - Thermal stress leading to population decline (2) | Adverse Outcome |
| Aop:324 - Thermal stress leading to population decline (1) | Adverse Outcome |
| Aop:363 - Thyroperoxidase inhibition leading to increased mortality via altered retinal layer structure | Adverse Outcome |
| Aop:349 - Inhibition of 11β-hydroxylase leading to decresed population trajectory | Adverse Outcome |
| Aop:348 - Inhibition of 11β-Hydroxysteroid Dehydrogenase leading to decreased population trajectory | Adverse Outcome |
| Aop:376 - Androgen receptor agonism leading to male-biased sex ratio | Adverse Outcome |
| Aop:386 - Increased reactive oxygen species production leading to population decline via inhibition of photosynthesis | Adverse Outcome |
| Aop:387 - Increased reactive oxygen species production leading to population decline via mitochondrial dysfunction | Adverse Outcome |
| Aop:388 - DNA damage leading to population decline via programmed cell death | Adverse Outcome |
| Aop:389 - Oxygen-evolving complex damage leading to population decline via inhibition of photosynthesis | Adverse Outcome |
| Aop:364 - Thyroperoxidase inhibition leading to increased mortality via decreased eye size | Adverse Outcome |
| Aop:365 - Thyroperoxidase inhibition leading to increased mortality via altered photoreceptor patterning | Adverse Outcome |
| Aop:399 - Inhibition of Fyna leading to increased mortality via decreased eye size (Microphthalmos) | Adverse Outcome |

Biological Context

Level of Biological Organization

Population

Domain of Applicability

Taxonomic Applicability

| Term | Scientific Term | Evidence | Links |
|-------------|-----------------|----------|-------------|
| all species | all species | High | <u>NCBI</u> |

Life Stage Applicability

| Life Stage | Evidence |
|-----------------|---------------|
| All life stages | Not Specified |

Sex Applicability

| Sex | Evidence |
|------------|---------------|
| Unspecific | Not Specified |

Consideration of population size and changes in population size over time is potentially relevant to all living organisms.

Key Event Description

Population ecology is the study of the sizes (and to some extent also the distribution) of plant and animal populations and of the processes, mainly biological in nature, that determine these sizes. As such, it provides an integrated measure of events occurring at lower levels of biological organization (biochemical, organismal, etc.). The population size in turn determines community and ecosystem structure. For fish, maintenance of sustainable fish and wildlife populations (i.e., adequate to ensure long-term delivery of valued ecosystem services) is an accepted regulatory goal upon which risk assessments and risk management decisions are based.

How it is Measured or Detected

Population trajectories, either hypothetical or site specific, can be estimated via population modeling based on measurements of vital rates or reasonable surrogates measured in laboratory studies. As an example, Miller and Ankley 2004 used measures of cumulative fecundity from laboratory studies with repeat spawning fish species to predict population-level consequences of continuous exposure.

Regulatory Significance of the AO

Maintenance of sustainable fish and wildlife populations (i.e., adequate to ensure long-term delivery of valued ecosystem services) is a widely accepted regulatory goal upon which risk assessments and risk management decisions are based.

40 |

References

Miller DH, Ankley GT. 2004. Modeling impacts on populations: fathead minnow (Pimephales promelas) exposure to the endocrine disruptor 17ß-trenbolone as a case study. Ecotoxicology and Environmental Safety 59: 1-9.

Appendix 2 - List of Key Event Relationships in the AOP

List of Adjacent Key Event Relationships

Relationship: 1037: Inhibition, Deiodinase 1 leads to Decreased, Triiodothyronine (T3)

AOPs Referencing Relationship

| AOP Name | Adjacency | Weight of Evidence | Quantitative Understanding |
|---|-----------|-----------------------|-------------------------------|
| Deiodinase 1 inhibition leading to increased mortality via reduced posterior swim bladder inflation | adjacent | Low | Low |
| Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation | adjacent | Low | Low |

Evidence Supporting Applicability of this Relationship

Taxonomic Applicability

| Term | Scientific Term | Evidence | Links |
|----------------|---------------------|----------|-------------|
| zebrafish | Danio rerio | Low | <u>NCBI</u> |
| fathead minnow | Pimephales promelas | Low | <u>NCBI</u> |

Life Stage Applicability

| Life Stage | Evidence |
|-----------------|----------|
| All life stages | High |

Sex Applicability

| Sex | Evidence |
|------------|----------|
| Unspecific | Moderate |

Taxonomic: Deiodinases are important for the activation of T4 to T3 across vertebrates. Therefore, this KER is plausibly applicable across vertebrates. There appear to be differences among vertebrate classes relative to the role of the different deiodinase isoforms in regulating thyroid hormone levels. It is generally assumed that deiodinase 1 in liver is the main supplier of T3 to circulation in mammals (Leonard et al., 1986), and the same appears to be true for birds (Freeman et al., 1991), while DIO2 is assumed to regulate intracellular concentrations of T3. In contrast to the general assumptions however, Maia et al. (2005) determined that in a normal physiological situation in humans the contribution of DIO2 to plasma T3 levels is twice that of DIO1. By contrast, DIO1 function in teleostean and amphibian T3 plasma regulation is less clear (Finnson et al. 1999, Kuiper et al. 2006). The presence of DIO1 in the liver of teleosts has been a controversial issue, and both the high level of DIO2 activity and its expression in the liver of teleosts are unique among vertebrates (Orozco and Valverde, 2005). These differences make it difficult to exactly evaluate the importance of DIO1

42 |

in regulating serum/tissue T3 levels across vertebrates. Mol et al. (1998) concluded that deiodinases in teleosts were more similar to mammalian deiodinases than had been generally accepted, based on the similarities in susceptibility to inhibition and the agreement of the Km values.

Life stage: Deiodinases are important for the activation of T4 to T3 across all life stages.

Sex: The KE is plausibly applicable to both sexes. Thyroid hormones are essential in both sexes and the components of the HPT-axis are identical in both sexes. There can however be sex-dependent differences in the sensitivity to the disruption of thyroid hormone levels and the magnitude of the response. In humans, females appear more susceptible to hypothyroidism compared to males when exposed to certain halogenated chemicals (Hernandez-Mariano et al., 2017; Webster et al., 2014). In adult zebrafish, Liu et al. (2019) showed sex-dependent changes in thyroid hormone levels and mRNA expression of regulatory genes including corticotropin releasing hormone (crh), thyroid stimulating hormone (tsh) and deiodinase 2 after exposure to organophosphate flame retardants. The underlying mechanism of any sex-related differences remains unclear.

Key Event Relationship Description

The two major thyroid hormones are thyroxine (T4) and the more biologically active triiodothyronine (T3), both iodinated derivatives of tyrosine. Active and inactive THs are tightly regulated by enzymes called iodothyronine deiodinases (DIO). The activation occurs via outer ring deiodination (ORD), i.e. removing iodine from the outer, phenolic ring of T4 to form T3, while inactivation occurs via inner ring deiodination (IRD), i.e. removing iodine from the inner tyrosol ring of T4 or T3.

Three types of iodothyronine deiodinases (DIO1-3) have been described in vertebrates that activate or inactivate THs and are therefore important mediators of TH action. All deiodinases are integral membrane proteins of the thioredoxin superfamily that contain selenocysteine in their catalytic centre. Type I deiodinase is capable of converting T4 into T3, as well as to convert rT3 to the inactive thyroid hormone 3,3' T2, through outer ring deiodination. rT3, rather than T4, is the preferred substrate for DIO1. furthermore, DIO1 has a very high Km (µM range, compared to nM range for DIO2) (Darras and Van Herck, 2012). Type II deiodinase (DIO2) is only capable of ORD activity with T4 as a preferred substrate (i.e., activation of T4 tot T3). DIO3 can inner ring deiodinate T4 and T3 to the inactive forms of THs, reverse T3, (rT3) and 3,3'-T2 respectively. (Darras and Van Herck, 2012)

Because of the high Km and preference for rT3 as a substrate, the importance of DIO1 in activating T4 to T3 in a physiological situation is likely limited.

Evidence Supporting this KER

Inhibition of DIO1 activity is widely accepted to directly decrease T3 levels, since the conversion of T4 to T3 is inhibited. The importance of DIO1 inhibition in altering serum and/or tissue T3 levels depends on the relative role of different deiodinases in regulating serum versus tissue T3 levels and in negative feedback within the HPT axis. Both aspects appear to differ to some extent among vertebrate taxa.

Biological Plausibility

Inhibition of DIO1 activity is widely accepted to directly decrease T3 levels, since the conversion of T4 to T3 is inhibited.

Empirical Evidence

- In the study of Cavallin et al. (2017) fathead minnow larvae were exposed to iopanoic acid, a model iodothyronine deiodinase inhibitor that is assumed to inhibit all three deiodinase enzymes (DIO1,2,3) and pronounced decreases of whole body T3 concentrations were observed.
- Stinckens et al. (2020) showed that iopanoic acid reduced whole-body T3 levels in zebrafish in 21 and 32 day old larvae that had been exposed starting from fertilization.
- Stinckens et al. (2018) showed that perfluorooctanoic acid (PFOA) is a DIO1 and DIO2 inhibitor, and Wang et al. (2020) showed that T3 levels were decreased in zebrafish exposed continuously until the age of 5 days 250 or 500 mg/L. They also showed a T4 decrease, which is unexpected upon exposure to a DIO inhibitor. This is possibly due to one or more additional thyroid hormone disruption mechanisms of PFOA.

Uncertainties and Inconsistencies

Since in fish early life stages THs are typically measured on a whole body level, it is currently uncertain whether T3 level changes occur at the serum and/or tissue level.

The importance of DIO1 inhibition in altering serum or tissue T3 levels depends on the relative role of different deiodinases in regulating serum versus tissue T3 levels and in negative feedback within the HPT axis. Both aspects appear to differ to some extent among vertebrate taxa, but the details are not understood yet.

Another uncertainty lies in the relative importance of the different T4 activating iodothyronine deiodinases (DIO1, DIO2) in the conversion of T4 to T3. It has been previously suggested that DIO2 is the major contributor to TH activation in developing zebrafish embryos (Darras et al., 2015; Walpita et al., 2010). It has been shown that a morpholino knockdown targeting DIO1 mRNA alone did not affect embryonic development in zebrafish, while knockdown of DIO2 delayed progression of otic vesicle length, head-trunk angle and pigmentation index (Houbrechts et al., 2016; Walpita et al., 2010, 2009). DIO1 inhibition may only become essential in hypothyroidal circumstances, for example when DIO2 is inhibited or in case of iodine deficiency, in zebrafish (Walpita et al., 2010) and mice (Galton et al., 2009; Schneider et al., 2006).

In the study of Cavallin et al. (2017) fathead minnow larvae were exposed to IOP, a model iodothyronine deiodinase inhibitor that is assumed to inhibit all three deiodinase enzymes (DIO1,2,3). Transcriptional analysis showed that especially DIO2, but also DIO3 mRNA levels (in some treatments), were increased in 10 to 21 day old larvae exposed to IOP as of the age of 6 days. This suggests that IOP effectively inhibited DIO2 and DIO3 in the larvae and that mRNA levels increased as a compensatory response. The authors also observed pronounced decreases of whole body T3 concentrations and increases of whole body T4 concentrations. It is not clear whether inhibition of DIO1 also played a role in the decrease of T3 levels.

Quantitative Understanding of the Linkage

Since in fish enzyme activity and thyroid hormone levels are rarely measured in the same study, quantitative understanding of this linkage is limited.

Known Feedforward/Feedback loops influencing this KER

Thyroid hormone levels are regulated via negative feedback, in part via regulation of the expression of all three DIO isoforms in response to deviating TH levels. This feedback mechanism influences this KER. Additionally, deiodinases regulate the activity of thyroid hormones, not only in serum and target organs, but also in the thyroid gland. On top of that, deiodinases themselves are known to be involved in the negative

feedback system that results in increased TSH levels when the levels of T4 (and also T3) in serum are low (Schneider et al., 2001), resulting in an even more complicated impact on this KER. Increased TSH levels then stimulate increased T4 release from the thyroid gland, resulting in a compensatory increase of serum T4 levels. In DIO2 knockout mice it seemed that the negative feedback system was blocked resulting in increased levels of T4 and TSH and in normal rather than decreased T3 levels compared to WT. By inhibiting DIO1 using a PTU exposure, Schneider et al. (2001) showed that DIO2 played a role in the increased TSH levels in response to T3 or T4 injection in mice.

References

Cavallin, J.E., Ankley, G.T., Blackwell, B.R., Blanksma, C.A., Fay, K.A., Jensen, K.M., Kahl, M.D., Knapen, D., Kosian, P.A., Poole, S.T., Randolph, E.C., Schroeder, A.L., Vergauwen, L., Villeneuve, D.L., 2017. Impaired swim bladder inflation in early life stage fathead minnows exposed to a deiodinase inhibitor, iopanoic acid. Environmental Toxicology and Chemistry 36, 2942-2952.

Darras, V.M., Van Herck, S.L.J., 2012. Iodothyronine deiodinase structure and function: from ascidians to humans. Journal of Endocrinology 215, 189-206.

Finnson, K.W., McLeese, J.M., Eales, J.G., 1999. Deiodination and deconjugation of thyroid hormone conjugates and type I deiodination in liver of rainbow trout, Oncorhynchus mykiss. General and Comparative Endocrinology 115, 387-397.

Hernandez-Mariano JA, Torres-Sanchez L, Bassol-Mayagoitia S, Escamilla-Nunez M, Cebrian ME, Villeda-Gutierrez EA, Lopez-Rodriguez G, Felix-Arellano EE, Blanco-Munoz J. 2017. Effect of exposure to p,p '-dde during the first half of pregnancy in the maternal thyroid profile of female residents in a mexican floriculture area. Environmental Research. 156:597-604.

Kuiper, G., Klootwijk, W., Dubois, G.M., Destree, O., Darras, V.M., Van der Geyten, S., Demeneix, B., Visser, T.J., 2006. Characterization of recombinant Xenopus laevis type I iodothyronine deiodinase: substitution of a proline residue in the catalytic center by serine (Pro132Ser) restores sensitivity to 6-propyl-2-thiouracil. Endocrinology 147, 3519-3529.

Liu XS, Cai Y, Wang Y, Xu SH, Ji K, Choi K. 2019. Effects of tris(1,3-dichloro-2-propyl) phosphate (tdcpp) and triphenyl phosphate (tpp) on sex-dependent alterations of thyroid hormones in adult zebrafish. Ecotoxicology and Environmental Safety. 170:25-32.

Mol, K.A., Van der Geyten, S., Burel, C., Kuhn, E.R., Boujard, T., Darras, V.M., 1998. Comparative study of iodothyronine outer ring and inner ring deiodinase activities in five teleostean fishes. Fish Physiology and Biochemistry 18, 253-266.

Schneider, M.J., Fiering, S.N., Pallud, S.E., Parlow, A.F., St Germain, D.L., Galton, V.A., 2001. Targeted disruption of the type 2 selenodeiodinase gene (D102) results in a phenotype of pituitary resistance to T-4. Molecular Endocrinology 15, 2137-2148.

Stinckens, E., Vergauwen, L., Ankley, G.T., Blust, R., Darras, V.M., Villeneuve, D.L., Witters, H., Volz, D.C., Knapen, D., 2018. An AOP-based alternative testing strategy to predict the impact of thyroid hormone disruption on swim bladder inflation in zebrafish. Aquatic Toxicology 200, 1-12.

Stinckens, E., Vergauwen, L., Blackwell, B.R., Anldey, G.T., Villeneuve, D.L., Knapen, D., 2020. Effect of Thyroperoxidase and Deiodinase Inhibition on Anterior Swim Bladder Inflation in the Zebrafish. Environmental Science & Technology 54, 6213-6223.

Wang, J.X., Shi, G.H., Yao, J.Z., Sheng, N., Cui, R.N., Su, Z.B., Guo, Y., Dai, J.Y., 2020. Perfluoropolyether carboxylic acids (novel alternatives to PFOA) impair zebrafish posterior swim bladder development via thyroid hormone disruption. Environment International 134.

Webster GM, Venners SA, Mattman A, Martin JW. 2014. Associations between perfluoroalkyl acids (pfass) and maternal thyroid hormones in early pregnancy: A population-based cohort study. Environmental Research. 133:338-347.

Relationship: 1035: Decreased, Triiodothyronine (T3) leads to Reduced, Anterior swim bladder inflation

AOPs Referencing Relationship

| AOP Name | Adjacency | Weight of Evidence | Quantitative Understanding |
|---|-----------|-----------------------|-------------------------------|
| Deiodinase 2 inhibition leading to increased mortality via reduced anterior swim bladder inflation | adjacent | Moderate | Moderate |
| Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation | adjacent | Moderate | Moderate |
| Thyroperoxidase inhibition leading to increased mortality via reduced anterior swim bladder inflation | adjacent | Moderate | Moderate |

Evidence Supporting Applicability of this Relationship

Taxonomic Applicability

| Term | Scientific Term | Evidence | Links |
|----------------|---------------------|----------|-------------|
| zebrafish | Danio rerio | Moderate | <u>NCBI</u> |
| fathead minnow | Pimephales promelas | High | NCBI |

Life Stage Applicability

| Life Stage | Evidence |
|------------|----------|
| Larvae | High |

Sex Applicability

| Sex | Evidence |
|------------|----------|
| Unspecific | Moderate |

Taxonomic: Teleost fish can be divided in two groups according to swim bladder morphology: physoclistous (e.g., yellow perch, sea bass, striped bass, medaka) and physostomous (e.g., zebrafish and fathead minnow). Physostomous fish retain a duct between the digestive tract and the swim bladder during adulthood allowing them to gulp air at the surface to fill the swim bladder. In contrast, in physoclistous fish, once initial inflation by gulping atmospheric air at the water surface has occurred, the swim bladder is closed off from the digestive tract and swim bladder volume is regulated by gas secretion into the swim bladder (Woolley and Qin, 2010). The evidence for impaired inflation of the anterior chamber of the swim bladder currently comes from work on zebrafish and fathead minnow (Stinckens et al., 2016; Nelson et al., 2016; Cavallin et al., 2017; Godfrey et al., 2017; Stinckens et al., 2020). While zebrafish and fathead minnows are physostomous fish with a two-chambered swim bladder, the Japanese rice fish or medaka (*Oryzias latipes*) is a physoclistous fish with a single chambered swim bladder that inflates during early development. This KER is not applicable to such fish species. Therefore, the current key event is plausibly applicable to physostomous fish in general.

Life stage: The anterior chamber inflates during a specific developmental time frame. In zebrafish, the anterior chamber inflates around 21 days post fertilization (dpf) which is during the larval stage. In the fathead minnow, the anterior chamber inflates around 14 dpf, also during the larval stage. Therefore this KER is only applicable to the larval life stage.

Sex: This KER plausibly applicable to both sexes. Sex differences are not often investigated in tests using early life stages of fish. For zebrafish and fathead minnow, it is currently unclear whether sex-related differences are important in determining the magnitude of the changes in this KER. Different fish species have different sex determination and differentiation strategies. Zebrafish do not have identifiable heteromorphic sex chromosomes and sex is determined by multiple genes and influenced by the environment (Nagabhushana and Mishra, 2016). Zebrafish are undifferentiated gonochorists since both sexes initially develop an immature ovary (Maack and Segner, 2003). Immature ovary development progresses until approximately the onset of the third week. Later, in female fish immature ovaries continue to develop further, while male fish undergo transformation of ovaries into testes. Final transformation into testes varies among male individuals, however finishes usually around 6 weeks post fertilization. Since the anterior chamber inflates around 21 days post fertilization in zebrafish, sex differences are expected to play a minor role. Fathead minnow gonad differentiation also occurs during larval development. Fathead minnows utilize a XY sex determination strategy and markers can be used to genotype sex in life stages where the sex is not yet clearly defined morphologically (Olmstead et al., 2011). Ovarian differentiation starts at 10 dph followed by rapid development (Van Aerle et al., 2004). At 25 dph germ cells of all stages up to the primary oocytes stage were present and at 120 dph, vitellogenic oocytes were present. The germ cells (spermatogonia) of the developing testes only entered meiosis around 90-120 dph. Mature testes with spermatozoa are present around 150 dph. Since the anterior chamber inflates around 14 days post fertilization (9 dph) in fathead minnows, sex differences are expected to play a minor role in the current KER.

Key Event Relationship Description

Thyroid hormones are known to be involved in development, especially in metamorphosis in amphibians and in embryonic-to-larval transition and larval-to-juvenile transition, including anterior chamber inflation in fish. Reduced T3 levels prohibit local TH action in the target tissues. Since swim bladder development and/or inflation is regulated by thyroid hormones, this results in impaired anterior chamber inflation.

Evidence Supporting this KER

There is convincing evidence that decreased T3 levels result in impaired anterior chamber inflation, but the underlying mechanisms are not completely understood. A very convincing linear quantitative relationship between reduced T3 levels and reduced anterior chamber volume was shown in zebrafish across exposure to a limited set of three compounds. Therefore the evidence supporting this KER can be considered moderate.

Biological Plausibility

Thyroid hormones are known to be involved in development, especially in metamorphosis in amphibians and in embryonic-to-larval transition (Liu and Chan, 2002) and larval-to-juvenile transition (Brown et al., 1997) in fish. Inflation of the anterior swim bladder chamber is part of the larval-to-juvenile transition in fish, together with the development of adult fins and fin rays, ossification of the axial skeleton, formation of an adult

pigmentation pattern, scale formation, maturation and remodelling of organs including the lateral line, nervous system, gut and kidneys (Brown, 1997; Liu and Chan, 2002; McMenamin and Parichy, 2013).

Empirical Evidence

Dedicated studies with two different experimental setups have been conducted to investigate the link between reduced T3 levels and reduced anterior chamber inflation:

1. Studies applying larval exposures initiated after posterior chamber inflation

- In a study in which larval fathead minnows (*Pimephales promelas*) were exposed to the thyroid peroxidase inhibitor 2-mercaptobenzothiazole (MBT), T3 concentrations measured at 14dpf were reduced at the same concentration (1 mg/L) that significantly reduced anterior swim bladder inflation at the same time-point (Nelson et al. 2016).
- In the study of Cavallin et al. (2017) fathead minnow larvae were exposed to IOP, a model iodothyronine deiodinase inhibitor that is assumed to inhibit all three deiodinase enzymes (DIO1,2,3). The authors observed pronounced decreases of whole body T3 concentrations and increases of whole body T4 concentrations, together with impaired inflation of the anterior swim bladder chamber. More specifically, inflation was delayed and the size of the swim bladder chamber was reduced until the end of the exposure experiment.

Since exposures were started after inflation of the posterior chamber, these studies show that DIO inhibition can directly affect anterior chamber inflation.

2. Studies applying continuous exposure initiated immediately after fertilization and thus including both posterior and anterior chamber inflation

In the study of Stinckens et al. (2020) exposure concentrations were chosen where the posterior chamber inflates. A strong correlation between reduced T3 levels and reduced anterior chamber inflation was observed in zebrafish exposed to iopanoic acid, a deiodinase inhibitor, as well as methimazole and propylthiouracil, both thyroperoxidase inhibitors, from fertilization until the age of 32 days. Anterior chamber inflation was delayed and a number of larvae did not manage to inflate the anterior chamber by the end of the 32 day exposure period. Additionally, exposed fish that had inflated the swim bladder had reduced anterior chamber sizes.

Uncertainties and Inconsistencies

- Since in fish early life stages THs are typically measured on a whole-body level, it is currently uncertain whether TH levels changes occur at the serum and/or tissue level.
- The mechanism underlying the link between reduced T3 and reduced anterior chamber inflation remains unclear, but several hypotheses exist (Stinckens et al., 2020). For example, altered gas distribution between chambers could be the result of impaired development of smooth muscle fibers, delayed and/or impaired evagination of the anterior chamber, impaired anterior budding through altered Wnt and hedgehog signalling, etc. Reinwald et al. (2021) showed that T3 and propylthiouracil treatment of zebrafish embryos altered expression of genes involved in muscle contraction and functioning in an opposing fashion. The authors suggested impaired muscle function as an additional key event between decreased T3 levels and reduced swim bladder inflation.
- Increased T3 levels also seem to result in reduced swim bladder inflation. For example, Li et al. (2011) reported impairment of swim bladder inflation in Chinese rare minnows (*Gobiocypris rarus*) exposed to exogenous T3.

References

Brown, D.D., 1997. The role of thyroid hormone in zebrafish and axolotl development. Proceedings of the National Academy of Sciences of the United States of America 94, 13011-13016.

Cavallin, J.E., Ankley, G.T., Blackwell, B.R., Blanksma, C.A., Fay, K.A., Jensen, K.M., Kahl, M.D., Knapen, D., Kosian, P.A., Poole, S.T., Randolph, E.C., Schroeder, A.L., Vergauwen, L., Villeneuve, D.L., 2017. Impaired swim bladder inflation in early life stage fathead minnows exposed to a deiodinase inhibitor, iopanoic acid. Environmental Toxicology and Chemistry 36, 2942-2952.

Godfrey, A., Hooser, B., Abdelmoneim, A., Horzmann, K.A., Freemanc, J.L., Sepulveda, M.S., 2017. Thyroid disrupting effects of halogenated and next generation chemicals on the swim bladder development of zebrafish. Aquatic Toxicology 193, 228-235.

Li W, Zha J, Yang L, Li Z, Wang Z. Regulation of thyroid hormone related genes mRNA expression by exogenous T_3 in larvae and adult Chinese rare minnow (Gobiocypris rarus). Environ Toxicol Pharmacol. 2011 Jan;31(1):189-97. doi: 10.1016/j.etap.2010.10.007.

Liu, Y.W., Chan, W.K., 2002. Thyroid hormones are important for embryonic to larval transitory phase in zebrafish. Differentiation 70, 36-45.

McMenamin, S.K., Parichy, D.M., 2013. Metamorphosis in Teleosts. Animal Metamorphosis 103, 127-165.

Nagabhushana A, Mishra RK. 2016. Finding clues to the riddle of sex determination in zebrafish. Journal of Biosciences. 41(1):145-155.

Nelson KR, Schroeder AL, Ankley GT, Blackwell BR, Blanksma C, Degitz SJ, Flynn KM, Jensen KM, Johnson RD, Kahl MD, Knapen D, Kosian PA, Milsk RY, Randolph EC, Saari T, Stinckens E, Vergauwen L, Villeneuve DL. Impaired anterior swim bladder inflation following exposure to the thyroid peroxidase inhibitor 2-mercaptobenzothiazole part I: Fathead minnow. Aquat Toxicol. 2016 Apr;173:192-203. doi: 10.1016/j.aquatox.2015.12.024.

Olmstead AW, Villeneuve DL, Ankley GT, Cavallin JE, Lindberg-Livingston A, Wehmas LC, Degitz SJ. 2011. A method for the determination of genetic sex in the fathead minnow, pimephales promelas, to support testing of endocrine-active chemicals. Environmental Science & Technology. 45(7):3090-3095.

Reinwald H, Konig A, Ayobahan SU, Alvincz J, Sipos L, Gockener B, Bohle G, Shomroni O, Hollert H, Salinas G et al. 2021. Toxicogenomic fin(ger)prints for thyroid disruption aop refinement and biomarker identification in zebrafish embryos. Science of the Total Environment. 760.

Stinckens, E., Vergauwen, L., Blackwell, B.R., Anldey, G.T., Villeneuve, D.L., Knapen, D., 2020. Effect of Thyroperoxidase and Deiodinase Inhibition on Anterior Swim Bladder Inflation in the Zebrafish. Environmental Science & Technology 54, 6213-6223.

Stinckens, E., Vergauwen, L., Schroeder, A., Maho, W., Blackwell, B., Witters, H., Blust, R., Ankley, G., Covaci, A., Villeneuve, D., Knapen, D., 2016. Impaired anterior swim bladder inflation following exposure to the thyroid peroxidase inhibitor 2-mercaptobenzothiazole part II: Zebrafish. Aquatic Toxicology 173, 204-217.

Uchida, D., Yamashita, M., Kitano, T., Iguchi, T., 2002. Oocyte apoptosis during the transition from ovarylike tissue to testes during sex differentiation of juvenile zebrafish. Journal of Experimental Biology 205, 711-718.

van Aerle R, Runnalls TJ, Tyler CR. 2004. Ontogeny of gonadal sex development relative to growth in fathead minnow. Journal of Fish Biology. 64(2):355-369.Zeng FX, Sherry JP, Bols NC. 2016. Evaluating the toxic potential of benzothiazoles with the rainbow trout cell lines, rtgill-w1 and rtl-w1. Chemosphere. 155:308-318.

50 |

Relationship: 1034: Reduced, Anterior swim bladder inflation leads to Reduced, Swimming performance

AOPs Referencing Relationship

| AOP Name | Adjacency | Weight of Evidence | Quantitative Understanding |
|---|-----------|-----------------------|-------------------------------|
| Deiodinase 2 inhibition leading to increased mortality via reduced anterior swim bladder inflation | adjacent | Moderate | Low |
| Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation | adjacent | Moderate | Low |
| Thyroperoxidase inhibition leading to increased mortality via reduced anterior swim bladder inflation | adjacent | Moderate | Low |

Evidence Supporting Applicability of this Relationship

Taxonomic Applicability

| Term | Scientific Term | Evidence | Links |
|----------------|---------------------|----------|-------------|
| zebrafish | Danio rerio | High | <u>NCBI</u> |
| fathead minnow | Pimephales promelas | Low | NCBI |

Life Stage Applicability

| Life Stage | Evidence |
|------------|----------|
| Larvae | High |

Sex Applicability

| Sex | Evidence |
|------------|----------|
| Unspecific | Moderate |

Taxonomic: Importance of proper functioning of the swim bladder for supporting natural swimming behaviour can be plausibly assumed to be generally applicable to fish possessing an anterior chamber. Evidence exists for the role of the posterior chamber in swimming performance comes from a wide variety of freshwater and marine fish species. Evidence for the specific role of the anterior chamber is however less abundant.

Life stage: In zebrafish, the anterior chamber inflates around 21 days post fertilization (dpf) which is during the larval stage. In the fathead minnow, the anterior chamber inflates around 14 dpf, also during the larval stage. Therefore this KER is only applicable to the larval life stage. To what extent fish can survive and swim with partly inflated swim bladders during later life stages is unknown.

Sex: This KER plausibly applicable to both sexes. Sex differences are not often investigated in tests using early life stages of fish. For zebrafish and fathead minnow, it is currently unclear whether sex-related differences are important in determining the magnitude of the changes in this KER. Different fish species have different sex determination and differentiation strategies. Zebrafish do not have identifiable

heteromorphic sex chromosomes and sex is determined by multiple genes and influenced by the environment (Nagabhushana and Mishra, 2016). Zebrafish are undifferentiated gonochorists since both sexes initially develop an immature ovary (Maack and Segner, 2003). Immature ovary development progresses until approximately the onset of the third week. Later, in female fish immature ovaries continue to develop further, while male fish undergo transformation of ovaries into testes. Final transformation into testes varies among male individuals, however finishes usually around 6 weeks post fertilization. Since the anterior chamber inflates around 21 days post fertilization in zebrafish, sex differences are expected to play a minor role. Fathead minnow gonad differentiation also occurs during larval development. Fathead minnows utilize a XY sex determination strategy and markers can be used to genotype sex in life stages where the sex is not yet clearly defined morphologically (Olmstead et al., 2011). Ovarian differentiation starts at 10 dph followed by rapid development (Van Aerle et al., 2004). At 25 dph germ cells of all stages up to the primary oocytes stage were present and at 120 dph, vitellogenic oocytes were present. The germ cells (spermatogonia) of the developing testes only entered meiosis around 90–120 dph. Mature testes with spermatozoa are present around 150 dph. Since the anterior chamber inflates around 14 days post fertilization (9 dph) in fathead minnows, sex differences are expected to play a minor role in the current KER.

Key Event Relationship Description

Effects on swim bladder inflation can alter swimming performance and buoyancy of fish, which is essential for predator avoidance, energy sparing, migration, reproduction and feeding behaviour, resulting in increased mortality.

Evidence Supporting this KER

The weight of evidence supporting a direct linkage between these two KEs, i.e. reduced anterior swim bladder inflation and reduced swimming performance, is weak.

Biological Plausibility

The anterior chamber of the swim bladder has a function in regulating the buoyancy of fish, by altering the volume of the swim bladder (Roberston et al., 2007). Fish rely on the lipid and gas content in their body to regulate their position within the water column, with the latter being more efficient at increasing body buoyancy. Therefore, fish with functional swim bladders have no problem supporting their body (Brix 2002), while it is highly likely that impaired inflation severely impacts swimming performance. Fish with no functional swim bladder can survive, but are severely disadvantaged, making the likelihood of surviving smaller.

Several studies in zebrafish and fathead minnow showed that a smaller AC was associated with a larger posterior chamber (Nelson et al., 2016; Stinckens et al., 2016; Cavallin et al., 2017, Stinckens et al., 2020) suggesting a possible compensatory mechanism. As shown by Stoyek et al. (2011) however, the AC volume is highly dynamic under normal conditions due to a series of regular corrugations running along the chamber wall, and is in fact the main driver for adjusting buoyancy while the basic PC volume remains largely invariable. Therefore, it is plausible to assume that functionality of the swim bladder is affected when AC inflation is incomplete, even when the PC appears to fully compensate the gas volume of the swim bladder.

Empirical Evidence

- Lindsey et al. (2010) showed that zebrafish started swimming deeper down in the water column upon inflation of the anterior chamber, confirming a role of the anterior chamber in supporting swimming performance.
- After exposure to 2-mercaptobenzothiazole, a TPO inhibitor, from 0 to 32 days post fertilization (dpf) in zebrafish, the swimming activity of fish was impacted starting at 26 dpf if the inflation of the anterior chamber of the swim bladder was impaired or had no normal structure/size (Stinckens et al., 2016).
- Methimazole (MMI) and propylthiouracil (PTU), two thyroperoxidase inhibitors, and iopanoic acid (IOP), a deiodinase inhibitor, each reduced both anterior chamber inflation and swimming distance in zebrafish exposed from fertlization until the age of 32 days (Stinckens et al., 2020). Stinckens et al. (2020) showed a specific, direct link between reduced anterior chamber inflation and reduced swimming performance.
 - First, after 21 d of exposure to 111 mg/L propylthiouracil around 30% of anterior chambers were not inflated and swimming distance was reduced, while by 32 days post fertilization all larvae had inflated their anterior chamber (although chamber surface was still smaller) and the effect on swimming distance had disappeared.
 - The most direct way to assess the role of anterior chamber inflation in swimming performance, however, is to compare larvae with and without inflated anterior chamber at the same time point and within the same experimental treatment. Both in the propylthiouracil exposure at 21 days post fertilization and in the iopanoic acid exposure at 21 and 32 days post fertilization, swimming distance was clearly reduced in larvae lacking an inflated anterior chamber, while the swimming distance of larvae with inflated anterior chamber was equal to that of controls.
 - Exposure concentrations were selected where the posterior chamber inflates. Even though the posterior chamber was generally larger when anterior chamber inflation was reduced, this did not remove the effect on swimming performance, confirming a direct link between proper anterior chamber inflation and swimming performance.
 - No morphological effects were observed, but in some treatments reduced length and/or condition factor was observed. However, reduced swimming performance after 32 days of IOP exposure to medium concentrations was not accompanied by reduced length or condition factor. Therefore, at least in this study no evidence was found that the effect on swimming performance was an indirect consequence of effects other than reduced swim bladder inflation.
- It has also been reported that larvae that fail to inflate their swim bladder use additional energy to maintain buoyancy (Lindsey et al., 2010, Goodsell et al. 1996), possibly contributing to reduced swimming activity.

Uncertainties and Inconsistencies

After exposure to 100 mg/L methimazole, 95% of the zebrafish larvae failed to inflate their anterior chamber at 32 dpf and swimming distance was reduced (Stinckens et al., 2020). On the other hand, there was no effect of impaired anterior chamber inflation on swimming distance in the methimazole exposure of 50 mg/L. Also, inflated but smaller anterior chambers did not result in a decreased swimming performance in this study. A similar result, where non-inflated anterior chambers did not consistently lead to reduced swimming performance, was previously found after exposure to 2-mercaptobenzothiazole (Stinckens et al., 2016). In summary, the precise relationship between these two KEs is not easy to determine and may be different for different chemicals. This is in part due to the complexity of the swim bladder system and the difficulty of distinguishing effects resulting from altered anterior chamber inflation from those resulting from altered posterior chamber inflation. Additionally, swimming capacity can be affected via other processes which may

or may not depend on the HPT axis, such as general malformations, decreased cardiorespiratory function, energy metabolism and growth.

As Robertson et al., (2007) reported, the swim bladder only starts regulating buoyancy actively from 32 dpf onward in zebrafish, possibly explaining the lack of effect on swimming capacity in some cases.

The anterior chamber is also important for producing and transducing sound through the Weberian Apparatus (Popper, 1974; Lechner and Ladich, 2008). It is highly plausible that impaired inflation or size of the anterior swim bladder could lead to increased mortality as hearing loss would affect their ability to respond to their surrounding environment, thus impacting ecological relevant endpoints such as predator avoidance or prey seeking (Wisenden et al., 2008; Fay, 2009).

References

Brix O (2002) The physiology of living in water. In: Hart PJ, Reynolds J (eds) Handbook of Fish Biology and Fisheries, Vol. 1, pp. 70–96. Blackwell Publishing, Malden, USA.

Cavallin, J.E., Ankley, G.T., Blackwell, B.R., Blanksma, C.A., Fay, K.A., Jensen, K.M., Kahl, M.D., Knapen, D., Kosian, P.A., Poole, S.T., Randolph, E.C., Schroeder, A.L., Vergauwen, L., Villeneuve, D.L., 2017. Impaired swim bladder inflation in early life stage fathead minnows exposed to a deiodinase inhibitor, iopanoic acid. Environmental Toxicology and Chemistry 36, 2942-2952.

Czesny, S.J., Graeb, B.D.S., Dettmersn, J.M., 2005. Ecological consequences of swimbladder noninflation for larval yellow perch. Trans. Am. Fish. Soc. 134,1011–1020, http://dx.doi.org/10.1577/T04-016.1.

Fay, R., 2009. Soundscapes and the sense of hearing of fishes. Integrative Zool. 4,26–32.

Goodsell, D.S., Morris, G.M., Olsen, A.J. 1996. Automated docking of fleixble ligands. Applications of Autodock. J. Mol. Recogonition, 9:1-5.

Lechner, W., Ladich, F., 2008. Size matters: diversity in swimbladders and Weberian ossicles affects hearing in catfishes. J. Exp. Biol. 211, 1681–1689.

Lindsey, B.W., Smith, F.M., Croll, R.P., 2010. From inflation to flotation: contribution of the swimbladder to whole-body density and swimming depth during development of the zebrafish (Danio rerio). Zebrafish 7, 85–96, http://dx.doi.org/10.1089/zeb.2009.0616.

Maack, G., Segner, H., 2003. Morphological development of the gonads in zebrafish. Journal of Fish Biology 62, 895-906.

Nagabhushana A, Mishra RK. 2016. Finding clues to the riddle of sex determination in zebrafish. Journal of Biosciences. 41(1):145-155.

Nelson, K., Schroeder, A., Ankley, G., Blackwell, B., Blanksma, C., Degitz, S., Flynn, K., Jensen, K., Johnson, R., Kahl, M., Knapen, D., Kosian, P., Milsk, R., Randolph, E., Saari, T., Stinckens, E., Vergauwen, L., Villeneuve, D., 2016. Impaired anterior swim bladder inflation following exposure to the thyroid peroxidase inhibitor 2-mercaptobenzothiazole part I: Fathead minnow. Aquatic Toxicology 173, 192-203.

Olmstead AW, Villeneuve DL, Ankley GT, Cavallin JE, Lindberg-Livingston A, Wehmas LC, Degitz SJ. 2011. A method for the determination of genetic sex in the fathead minnow, pimephales promelas, to support testing of endocrine-active chemicals. Environmental Science & Technology. 45(7):3090-3095.

Roberston, G.N., McGee, C.A.S., Dumbarton, T.C., Croll, R.P., Smith, F.M., 2007. Development of the swim bladder and its innervation in the zebrafish, Danio rerio. J. Morphol. 268, 967–985, http://dx.doi.org/10.1002/jmor.

Stinckens, E., Vergauwen, L., Blackwell, B.R., Anldey, G.T., Villeneuve, D.L., Knapen, D., 2020. Effect of Thyroperoxidase and Deiodinase Inhibition on Anterior Swim Bladder Inflation in the Zebrafish. Environmental Science & Technology 54, 6213-6223.

Stinckens, E., Vergauwen, L., Schroeder, A.L., Maho, W., Blackwell, B., Witter, H., Blust, R., Ankley, G.T., Covaci, A., Villenueve, D.L., Knapen, D., 2016. Disruption of thyroid hormone balance after 2-mercaptobenzothiazole exposure causes swim bladder inflation impairment—part II: zebrafish. Aquat. Toxicol. 173:204-17.

Stoyek, M.R., Smith, F.M., Croll, R.P., 2011. Effects of altered ambient pressure on the volume and distribution of gas within the swimbladder of the adult zebrafish, Danio rerio. Journal of Experimental Biology 214, 2962-2972.

van Aerle R, Runnalls TJ, Tyler CR. 2004. Ontogeny of gonadal sex development relative to growth in fathead minnow. Journal of Fish Biology. 64(2):355-369.

Wisenden, B.D., Pogatschnik, J., Gibson, D., Bonacci, L., Schumacher, A., Willet, A., 2008. Sound the alarm: learned association of predation risk with novelauditory stimuli by fathead minnows (Pimephales promelas) and glowlighttetras (Hemigrammus erythrozonus) after single simultaneous pairings withconspecific chemical alarm cues. Environ. Biol. Fish 81, 141–147.

Woolley, L.D., Qin, J.G., 2010. Swimbladder inflation and its implication to theculture of marine finfish larvae. Rev. Aquac. 2, 181–190, http://dx.doi.org/10.1111/j.1753-5131.2010.01035.x.

Relationship: 2212: Reduced, Swimming performance leads to Increased Mortality

AOPs Referencing Relationship

| AOP Name | Adjacency | Weight of evidence | Quantitative Understanding |
|---|-----------|--------------------|-------------------------------|
| Deiodinase 2 inhibition leading to increased mortality via reduced posterior swim bladder inflation | adjacent | Moderate | Low |
| Deiodinase 2 inhibition leading to increased mortality via reduced anterior swim bladder inflation | adjacent | Moderate | Low |
| Deiodinase 1 inhibition leading to increased mortality via reduced posterior swim bladder inflation | adjacent | Moderate | Low |
| Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation | adjacent | Moderate | Low |
| Thyroperoxidase inhibition leading to increased mortality via reduced anterior swim bladder inflation | adjacent | Moderate | Low |

Evidence Supporting Applicability of this Relationship

Taxonomic Applicability

| Term | Scientific Term | Evidence | Links |
|----------------|---------------------|----------|-------------|
| zebrafish | Danio rerio | Moderate | <u>NCBI</u> |
| fathead minnow | Pimephales promelas | Moderate | NCBI |

Life Stage Applicability

| Sex Applicability | | |
|-------------------|----------|--|
| Larvae | Moderate | |
| Juvenile | Moderate | |
| Adult | Moderate | |
| Life Stage | Evidence | |

| Sex | Evidence |
|------------|----------|
| Unspecific | Moderate |

Importance of swimming performance on survival is generally applicable to all hatched fish across life stages and sexes and to other taxa that rely on swimming to support vital behaviours.

Key Event Relationship Description

Reduced swimming performance is likely to affect essential endpoints such as predator avoidance, feeding behaviour and reproduction in taxa that rely on swimming to support these vital behaviours. These parameters are biologically plausible to affect survival, especially in a non-laboratory environment where food is scarce and predators are abundant.

Evidence Supporting this KER

A direct relationship between reduced swimming performance and reduced survival is difficult to establish. There is however a lot of indirect evidence linking reduced swim bladder inflation to reduced survival (https://aopwiki.org/relationships/2213), which can be plausibly assumed to be related to reduced swimming performance.

For example, all zebrafish larvae that failed to inflate the posterior chamber after exposure to 2 mg/L iopanoic acid (IOP), died by the age of 9 dpf (Stinckens et al., 2020). Since larvae from the same group that were able to inflate the posterior chamber survived and the test was performed in the laboratory in optimal conditions, it is plausible to assume that the cause of death was the inability to swim and find food due to the failure to inflate the posterior swim bladder chamber.

Biological Plausibility

Reduced swimming performance is likely to affect essential endpoints such as predator avoidance, feeding behaviour and reproduction. These parameters are biologically plausible to affect survival, especially in a non-laboratory environment where food is scarce and predators are abundant.

Empirical Evidence

A direct relationship between reduced swimming performance and reduced survival is difficult to establish. There is however a lot of indirect evidence linking reduced swim bladder inflation to reduced survival (see non-adjacent KER 1041), which can be plausibly assumed to be related to reduced swimming performance.

For example, all zebrafish larvae that failed to inflate the posterior chamber after exposure to 2 mg/L iopanoic acid (IOP), died by the age of 9 dpf (Stinckens et al., 2020). Since larvae from the same group that were able to inflate the posterior chamber survived and the test was performed in the laboratory in optimal conditions, it is plausible to assume that the cause of death was the inability to swim and find food due to the failure to inflate the posterior swim bladder chamber.

Uncertainties and Inconsistencies

A direct relationship between reduced swimming performance and reduced survival is difficult to establish in a laboratory environment where food is abundant and there are no predators.

Quantitative Understanding of the Linkage

Quantitative understanding of this linkage is currently limited.

Time-scale

Reduced swimming performance is not expected to immediately lead to mortality. Depending on the extent of the reduction in swimming performance and depending on the cause of death (e.g., starvation due to the inability to find food, being caught by a predator) the lag time may vary.

As an example, Stinckens et al. (2020) found that zebrafish larvae that failed to inflate the swim bladder at 5 dpf and did not manage to inflate it during the days afterwards died by the age of 9 dpf. Since zebrafish initiate exogenous feeding around 5 dpf when the yolk is almost completely depleted, there was a lag period of around 4 days after which reduced feeding resulted in mortality. Obviously, in a laboratory setup there is no increased risk of being caught by a predator.

References

Stinckens, E., Vergauwen, L., Blackwell, B.R., Anldey, G.T., Villeneuve, D.L., Knapen, D., 2020. Effect of Thyroperoxidase and Deiodinase Inhibition on Anterior Swim Bladder Inflation in the Zebrafish. Environmental Science & Technology 54, 6213-6223.

Relationship: 2013: Increased Mortality leads to Decrease, Population growth rate

AOPs Referencing Relationship

| AOP Name | Adjacency | Weight of evidence | Quantitative Understanding |
|--|-----------|--------------------|-------------------------------|
| Acetylcholinesterase Inhibition leading to Acute Mortality via Impaired Coordination & Movement | adjacent | | |
| Acetylcholinesterase inhibition leading to acute mortality | adjacent | Moderate | Moderate |
| Deiodinase 2 inhibition leading to increased mortality via reduced posterior swim bladder inflation | adjacent | Moderate | Moderate |
| Deiodinase 2 inhibition leading to increased mortality via reduced anterior swim bladder inflation | adjacent | Moderate | Moderate |
| Deiodinase 1 inhibition leading to increased mortality via reduced posterior swim bladder inflation | adjacent | Moderate | Moderate |
| Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation | adjacent | Moderate | Moderate |
| Thyroperoxidase inhibition leading to increased mortality via reduced anterior swim bladder inflation | adjacent | Moderate | Moderate |
| <u>Thyroperoxidase inhibition leading to</u> altered visual function via altered retinal layer structure | adjacent | Moderate | Moderate |
| Thyroperoxidase inhibition leading to altered visual function via decreased eye size | adjacent | | |
| Thyroperoxidaseinhibitionleadingtoalteredvisualfunctionviaalteredphotoreceptorpatterning | adjacent | | |
| Inhibition of Fyna leading to increased mortality via decreased eye size (Microphthalmos) | adjacent | High | High |
| GSK3beta inactivation leading to increased mortality via defects in developing inner ear | adjacent | High | High |

Evidence Supporting Applicability of this Relationship

Taxonomic Applicability

| Term | Scientific Term | Evidence | Links |
|----------------|---------------------|----------|-------------|
| zebrafish | Danio rerio | High | NCBI |
| fathead minnow | Pimephales promelas | High | <u>NCBI</u> |

Life Stage Applicability

| Life Stage | Evidence | |
|-------------------|----------|--|
| All life stages | High | |
| | | |
| Sex Applicability | | |
| Sex Applicability | Evidence | |

Taxonomic: All organisms must survive to reproductive age in order to reproduce and sustain populations. The additional considerations related to survival made above are applicable to other fish species in addition to zebrafish and fathead minnows with the same reproductive strategy (r-strategist as described in the theory of MaxArthur and Wilson (1967). The impact of reduced survival on population size is even greater for k-strategists that invest more energy in a lower number of offspring.

Life stage: Density dependent effects start to play a role in the larval stage of fish when free-feeding starts (Hazlerigg et al., 2014).

Sex: This linkage is independent of sex.

Key Event Relationship Description

Increased mortality in the reproductive population may lead to a declining population. This depends on the excess mortality due to the applied stressor and the environmental parameters such as food availability and predation rate. Most fish species are r- strategist, meaning they produce a lot of offspring instead of investing in parental care. This results in natural high larval mortality causing only a small percentage of the larvae to survive to maturity. If the excess larval mortality due to a stressor is small, the population dynamics might result in constant population size. Should the larval excess be more significant, or last on the long-term, this will affect the population. To calculate the long-term persistence of the population, population dynamic models should be used.

Evidence Supporting this KER

Survival rate is an obvious determinant of population size and is therefore included in population modeling (e.g., Miller et al., 2020).

Biological Plausibility

Survival to reproductive maturity is a parameter of demographic significance. Assuming resource availability (i.e., food, habitat, etc.) is not limiting to the extant population, sufficient mortality in the reproductive population may ultimately lead to declining population trajectories.

Under some conditions, reduced larval survival may be compensated by reduced predation and increased food availability, and therefore not result in population decline (Stige et al., 2019).

Empirical Evidence

According to empirical data, combined with population dynamic models, feeding larvae are the crucial life stage in zebrafish (and other r-strategists) for the regulation of the population. (Schäfers et al., 1993)

In fathead minnow, natural survival of early life stages has been found to be highly variable and influential on population growth (Miller and Ankley, 2004)

Rearick et al. (2018) used linked data from behavioural assays to survival trials and applied a modelling approach to quantify changes in antipredator escape performance of larval fathead minnows in order to predict changes in population abundance. This work was done in the context of exposure to an environmental oestrogen. Expsoed fish had delayed response times and slower escape speeds, and were more susceptible to predation. Population modelling showed that his can result in population decline.

In the context of fishing and fisheries, ample evidence of a link between increased mortality and a decrerase of population size has been given. Important insights can result from the investigation of optimum modes of fishing that allow for maintaining a population (Alekseeva and Rudenko, 2018). Jacobsen and Essington (2018) showed the impact of varying predation mortality on forage fish populations.

Boreman (1997) reviewed methods for comparing the population-level effects of mortality in fish populations induced by pollution or fishing.

Uncertainties and Inconsistencies

The extent to which larval mortality affects population size could depend on the fraction of surplus mortality compared to a natural situation.

There are scenarios in which individual mortality may not lead to declining population size. These include instances where populations are limited by the availability of habitat and food resources, which can be replenished through immigration. Effects of mortality in the larvae can be compensated by reduced competition for resources (Stige et al., 2019).

The direct impact of pesticides on migration behavior can be difficult to track in the field, and documentation of mortality during migration is likely underestimated (Eng 2017).

References

Alekseeva SM, Rudenko AI. 2018. Modeling of optimum fishing population. Marine Intellectual Technologies. 3(4):142-146.

Beaudouin, R., Goussen, B., Piccini, B., Augustine, S., Devillers, J., Brion, F., Pery, A.R., 2015. An individualbased model of zebrafish population dynamics accounting for energy dynamics. PloS one 10, e0125841.

Boreman J. 1997. Methods for comparing the impacts of pollution and fishing on fish populations. Transactions of the American Fisheries Society. 126(3):506-513.

Caswell, H., 2000. Matrix population models. Sinauer Sunderland, MA, USA.

Eng, M.L., Stutchbury, B.J.M. & Morrissey, C.A. Imidacloprid and chlorpyrifos insecticides impair migratory ability in a seed-eating songbird. Sci Rep 7, 15176 (2017)

Hazlerigg, C.R., Lorenzen, K., Thorbek, P., Wheeler, J.R., Tyler, C.R., 2012. Density-dependent processes in the life history of fishes: evidence from laboratory populations of zebrafish Danio rerio. PLoS One 7, e37550.

Jacobsen NS, Essington TE. 2018. Natural mortality augments population fluctuations of forage fish. Fish and Fisheries. 19(5):791-797.

MacArthur, R., Wilson, E., 1967. The Theory of Island Biogeography. Princeton: Princeton Univ. Press. 203 p.

Miller, D.H., Ankley, G.T., 2004. Modeling impacts on populations: fathead minnow (Pimephales promelas) exposure to the endocrine disruptor 17β -trenbolone as a case study. Ecotoxicology and Environmental Safety 59, 1-9.

Miller, D.H., Clark, B.W., Nacci, D.E. 2020. A multidimensional density dependent matrix population model for assessing risk of stressors to fish populations. Ecotoxicology and environmental safety 201, 110786.

Pinceel, T., Vanschoenwinkel, B., Brendonck, L., Buschke, F., 2016. Modelling the sensitivity of life history traits to climate change in a temporary pool crustacean. Scientific reports 6, 29451.

Rearick, D.C., Ward, J., Venturelli, P., Schoenfuss, H., 2018. Environmental oestrogens cause predationinduced population decline in a freshwater fish. Royal Society open science 5, 181065.

Schäfers, C., Oertel, D., Nagel, R., 1993. Effects of 3, 4-dichloroaniline on fish populations with differing strategies of reproduction. Ecotoxicology and Ecophysiology, 133-146.

Stige, L.C., Rogers, L.A., Neuheimer, A.B., Hunsicker, M.E., Yaragina, N.A., Ottersen, G., Ciannelli, L., Langangen, Ø., Durant, J.M., 2019. Density-and size-dependent mortality in fish early life stages. Fish and Fisheries 20, 962-976.

Hazlerigg, C.R.E., Tyler, C.R., Lorenzen, K., Wheeler, J.R., Thorbek, P., 2014. Population relevance of toxicant mediated changes in sex ratio in fish: An assessment using an individual-based zebrafish (Danio rerio) model. Ecological Modelling 280, 76-88.

Stige, L.C., Rogers, L.A., Neuheimer, A.B., Hunsicker, M.E., Yaragina, N.A., Ottersen, G., Ciannelli, L., Langangen, O., Durant, J.M., 2019. Density- and size-dependent mortality in fish early life stages. Fish and Fisheries 20, 962-976.

Annex 1: Weight of evidence evaluation table

AOP 158: Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation - Weight of evidence evaluation

| | Defining | High (Strong) | Moderate | Low (Weak) | | | |
|--|---|--|--|---|--|--|--|
| 1. Support for Biological Plausibility of KERs | Question Is there a mechanistic relationship between KE _{up} and KE _{down} consistent with established biological knowledge? | KER is plausible based on analogy to accepted biological relationships, but scientific understanding is incomplete | Empirical support for association between KEs , but the structural or functional relationship between them is not understood. | | | | |
| Relationship 1037: Inhibition, Deiodinase 1 (KE 1009) leads to Decreased, Triiodothyronine (T3) in serum (KE 1003) | Low DIO1 is capable of conve physiological situation is further depends on the levels and in negative fe typically measured on a the serum and/or tissue proxy for serum TH leve remains uncertain. | erting T4 to the more biolog s likely limited. The importa relative role of different deie edback within the HPT axis. whole body level, it is curre level. Pending more dedica els. In summary, there is a pl | nce of DIO1 inhibition in al odinases in regulating serur Finally, since in fish early l ently uncertain whether T3 ted studies, whole body TH | tering serum T3 levels n versus tissue T3 ife stages THs are level changes occur at levels are considered a | | | |
| Relationship 1035: Decreased, Triiodothyronine (T3) in serum (KE 1003) leads to Reduced, Anterior swim bladder inflation (KE 1007) | development, especially larval-to-juvenile transit to-juvenile transition in the axial skeleton, format remodeling of organs in empirical evidence, it is | cially the more biologically in metamorphosis in amph tion in fish. Inflation of the a fish, together with the deve titon of an adult pigmentatio cluding the lateral line, nerv plausible to assume that an understanding is incomplet | ibians and in embryonic-to interior swim bladder cham lopment of adult fins and fi on pattern, scale formation, rous system, gut and kidney terior inflation is under thy | -larval transition and aber is part of the larval- n rays, ossification of maturation and s. Together with | | | |
| Relationship 1034: Reduced, Anterior swim bladder inflation (KE 1007) leads to Reduced, Swimming performance (KE 1005) | buoyancy of fish. Stoyek under normal condition in fact the main driver fo largely invariable. There when anterior chamber compensate the gas volu | , the anterior chamber of th et al. (2011) showed that th s due to a series of regular c or adjusting buoyancy while fore, it is plausible to assun inflation is incomplete, even ume of the swim bladder. | ne anterior chamber volume orrugations running along the basic posterior chambe ne that functionality of the s | e is highly dynamic the chamber wall, and is er volume remains wim bladder is affected | | | |
| Relationship 2212: Reduced, Swimming performance (KE 1005) leads to Increaed mortality (KE 351) | feeding behaviour and r Apart from some indired laboratory. It may only h predators are abundant | formance is likely to affect e eproduction. These parame ct evidence, it has been diffio become apparent in a non-la | ters are biologically plausib cult to clearly establish this | le to affect survival. relationship in the | | | |
| Relationship 2013: Increased mortality (KE 351) leads to Decrease, Population trajectory (KE 360) | High It is widely accepted tha | t mortality increases, the po | opulation trajectory will eve | entually decrease. | | | |

AOP 158: Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation - Weight of evidence evaluation

| 2. Essentiality of KEs | Defining question | High (Strong) | Moderate | Low (Weak) |
|---|--|--|--|--|
| | Are downstream KEs | Direct evidence from | Indirect evidence that | No or contradictory |
| | and/or the AO prevented if an | specifically designed experimental studies | sufficient modification of an expected | experimental evidence of the essentiality of |
| | upstream KE is | illustrating essentiality | modulating factor | any of the KEs. |
| | blocked? | for at least one of the | attenuates or | |
| | | important KEs | augments a KE | |
| | | | | |
| KE 1009 (MIE): Inhibition, deiodinase | | | eported that knockdown of bladder chamber. Walpita | |
| 1 | reported reduced pigmer Dio2 knockdown fish. Th effects. These effects wer confirming the important 2010). | ntation, otic vesicle length is suggests that DIO1 is les e rescued after T3 supplen ce of T4 to T3 conversion b | and head-trunk angle in th is important than DIO2 in c nentation but not after T4 by Dio2 and perhaps also D | e same Dio1+2 and also causing downstream supplementation, bio1 (Walpita et al., 2009, |
| KE 1003: Decreased triiodothyronine (T3) in serum | reduced posterior chamb system in general. | er inflation, confirming a c | of decreased T3 levels for direct link between T3 leve | |
| | TH transporter MCT beta (Marelli et al., 2 zebrafish resulted in (2016) additionally with partially resist Walpita et al. (2009 angle in the same Di supplementation, but the assessed endpoid causing downstream (2) from chemical expose. Wang et al. (2020) c inflation in zebrafish and exogeneous T3 Maternal injection of to significant increate et al. (2019) showed larvae. Less information is avai inflation. Chopra et al. (2019) expected to lead to a synthesis - reduced Proving essentiality of r complicated by the com resulting from altered a | dinase 1 and 2 (Bagci et al., '8 (de Vrieze et al., 2014), 1 2016), and permanent known inpaired inflation of the showed that high T3 doses ant thyroid hormone recep , 2010) reported reduced p o1+2 and also Dio2 knock at not after T4 supplement nts in this study, this gene n effects upon disruption of the study, this gene the study, this gene the study, this gene n effects upon disruption of the study, this gene the study, the study the study the study the study play the study the study the study play the study the study the study the study play the study th | bigmentation, otic vesicle l down fish. These effects w ration. While swim bladder rally confirms the essentia of DIO1 and 2 function (Wa ble-body T3 as well as imp inoic acid and perfluoropo rtly rescued this effect. I T3 concentrations in the der inflation (Brown et al., increased posterior chaml of reduced T3 levels for r re showing that knockdown ince dual oxidase is impor | mone receptor alpha or ubrechts et al., 2016) in amber. Marelli et al. ative impact in mutants ength and head-trunk ere rescued after T3 inflation was not among lity of decreased T3 in lpita et al., 2009, 2010). aired posterior chamber lyether carboxylic acids eggs of striped bass lead 1988). Similarly, Molla ber diameter in zebrafish educed anterior chamber n of dual oxidase - tant for thyroid hormone ion is further of distinguishing effects |
| KE 1007: Reduced, anterior swim bladder inflation | chamber, larvae exposed reduced swimming dista | to PTU have a lower frequ nce. Later during the expos | at where control zebrafish ency of inflated anterior cl sure the frequency of non- stance disappeared confiru | nambers together with inflated anterior |
| | reduced anterior chambe | r inflation for the downstr | eam effect on swimming p | |
| KE 1005: Reduced, swimming performance | | this KE is difficult to achie | | |
| KE 351: Increased mortality | - | nortality is essential for re | duced population size. | |
| AOP as a whole | specifically designed exp combined DIO1 and DIO2 chamber inflation, but an evidence that reduced th Chopra et al. (2019) show synthesis, reduced anteri role in oxidative stress. T independent of DIO2 inhi | erimental studies illustrati 2 knockdown studies in zel terior chamber inflation w yroid hormone synthesis c ved that knockdown of dua or swim bladder inflation. here is no specific evidenc bition and DIO2 seems mo | w since there is limited dir ng essentiality. This incluc orafish showing the link wi vas not studied. There is ad auses reduced anterior sw al oxidase, important for th It should be noted that du e for the essentiality of DIO ore important than DIO1 in erall evidence for essentia | les evidence from ith reduced posterior Iditional indirect im bladder inflation: nyroid hormone al oxidase also plays a D1 inhibition providing sufficient T3 |

AOP 158: Deiodinase 1 inhibition leading to increased mortality via reduced anterior swim bladder inflation - Weight of evidence evaluation

| | Defining Questions | High (Strong) | Moderate | Low (Weak) | | | | | |
|--|--|--|---|--|--|--|--|--|--|
| 3. Empirical Support for KERs | Questions Does empirical evidence support that a change in KEup leads to an appropriate change in KEdown? Does KEup occur at lower doses and earlier time points than KE down and is the incidence of KEup > than that for KEdown? Inconsistencies? | if there is dependent change in both events following exposure to a wide range of specific stressors (extensive evidence for temporal, dose- response and incidence concordance) and no or few data gaps or conflicting data | if there is demonstrated dependent change in both events following exposure to a small number of specific stressors and some evidence inconsistent with the expected pattern that can be explained by factors such as experimental design, technical considerations, differences among laboratories, etc. | if there are limited or no studies reporting dependent change in both events following exposure to a specific stressor (i.e., endpoints never measured in the same study or not at all), and/or lacking evidence of temporal or dose- response concordance, or identification of significant inconsistencies in empirical support across taxa and species that don't align with the expected pattern for the hypothesised AOP | | | | | |
| Relationship 1037: Inhibition, Deiodinase 1 (KE 1009) leads to Decreased, Triiodothyronine (T3) in serum (KE 1003) | studies have shown that | ements of both KEs in the sa c chemicals able to inhibit D sus DIO2 is uncertain, and a | IO1 in vitro, reduce T3 leve | ls. The relative | | | | | |
| Relationship 1035: Decreased, Triiodothyronine (T3) in serum (KE 1003) leads to Reduced, Anterior swim bladder inflation (KE 1007) | evidence of dose concor TH levels result in impa | both T3 decreases and redu dance. Uncertainties mainly ired posterior chamber infla ition can only occur at a spe | relate to the mechanism thation. Temporal concordance | nrough which altered | | | | | |
| Relationship 1034: Reduced, Anterior swim bladder inflation (KE 1007) leads to Reduced, Swimming performance (KE 1005) Relationship 2212: Reduced, Swimming performance (KE | swimming performance specifically supported b propylthiouracil around reduced, while by 32 da chamber surface was sti Low A direct relationship bet | nce of a link between reduc including some evidence of y the study of Stinckens et a 30% of anterior chambers ys post fertilization all larva ll smaller) and the effect on tween reduced swimming p ere is however a lot of indire | dose concordance. Tempo l. (2020): First, after 21 d o were not inflated and swim had inflated their anterio swimming distance had dir erformance and increased i | ral concordance is f exposure to 111 mg/L uning distance was r chamber (although sappeared. mortality has been | | | | | |
| 1005) leads to Increaed mortality (KE 351) | inflation to increased ma related to reduced swim | ortality (see non-adjacent K | | | | | | | |
| Relationship 2013: Increased mortality (KE 351) leads to Decrease, Population trajectory (KE 360) | modeling. The extent to environmental exposure | us determinant of populatic which increased mortality r e scenario depends on the ci ompensated by reduced pre opulation decline. | nay impact population size ircumstances. Under some o | s in a realistic, conditions, reduced | | | | | |

| | | | | | | | dose and temporal conco | rdance | | | | | | | | | uncertainties, in | consistencies | | |
|-------------------------|----------------|-------------------------------|-----------------------|----------------------|------------------|---|-------------------------|--------------------|-----------------------|------------------------------|---|---------------------------------|---|--|---------------------------------|-------------------------|-----------------------|------------------------|---------------------------------|------------------------------|
| reference | species | chemical | expected MIE | exposure period | time point | concentrations tested | TPO inhibition | DIO1 inhibition | DIO2 inhibition | TH synthesis decreased | T4 in serum decreased | T3 in serum decreased | posterior swim bladder chamber inflation reduced | anterior swim bladder chamber inflation reduced | swimming performance reduced | increased mortality | decreased tpo mRNA | decreased dio1 mRNA | serum T4 increased | serum T3 increased |
| Cavallin et al. (2017) | fathead minnow | iopanoic acid | DIO1 and 2 inhibition | 0-6dpf | 4 dpf | 0.6, 1.9, 6.0 mg/L | n/a | n/a | n/a | n/a | n/a | .ť | n/a | n/a | n/a | | | | 0.6, 1.9, 6.0 mg/L ^f | 6 mg/L [£] |
| | fathead minnow | iopanoic acid | DIO1 and 2 inhibition | 0-6dpf | 6 dpf | 0.6, 1.9, 6.0 mg/L | n/a | | | n/a | n/a | . ⁴ | 6 mg/L | n/a | n/a | | | | ,t | 1.9, 6.0 mg/L ⁴ |
| Cavallin et al. (2017) | fathead minnow | iopanoic acid | DIO1 and 2 inhibition | 6-21 dpf | 10 dpf | 0.6, 1.9, 6.0 mg/L | n/a | | | n/a | n/a | 0.6, 1.9, 6.0 mg/L ^c | n/a | n/a | n/a | | | | 0.6, 1.9, 6.0 mg/L ^c | .t |
| Cavallin et al. (2017) | fathead minnow | iopanoic acid | DIO1 and 2 inhibition | 6-21 dpf | 14 dpf | 0.6, 1.9, 6.0 mg/L | n/a | | 0.6, 1.9, 6.0 mg/L* | n/a | n/a | 0.6, 1.9, 6.0 mg/L ^E | n/a | 0.6, 1.9, 6.0 mg/L | n/a | | | | 1.9, 6.0 mg/L ^f | .t |
| Cavallin et al. (2017) | fathead minnow | iopanoic acid | DIO1 and 2 inhibition | 6-21 dpf | 18 dpf | 0.6, 1.9, 6.0 mg/L | n/a | | 0.6, 1.9, 6.0 mg/L* | n/a | n/a | 0.6, 1.9, 6.0 mg/L ^c | n/a | 0.6, 1.9, 6.0 mg/L | n/a | | | | 0.6, 1.9, 6.0 mg/L ² | ,t |
| Cavallin et al. (2017) | fathead minnow | iopanoic acid | DIO1 and 2 inhibition | 6-21 dpf | 21 dpf | 0.6, 1.9, 6.0 mg/L | n/a | | 0.6, 1.9, 6.0 mg/L* | n/a | n/a | 0.6, 1.9, 6.0 mg/L ^r | n/a | 0.6, 1.9, 6.0 mg/L | n/a | 6 mg/L | | | 0.6, 1.9, 6.0 mg/L ² | .t |
| | | | | | | | | | 010, 210, 010 118, 2 | | ,= | | | | ., | | | | , ., | |
| Stinckens et al. (2016) | zebrafish | 2-mercaptobenzothiazole | TPO inhibition | 0-168 hpf | 120 hpf | 0.1, 0.35, 0.56, 0.7, 0.88, 1.75, 3.5, 7 mg/L | n/a | n/a | n/a | n/a | 0.35, 0.7 mg/L [£] (0.1 mg/L n | 0 | | n/a | 0.35, 0.56, 0.7, 0.88, 1.75, | 2 2 E 7 mali | | | t. | |
| Stinckens et al. (2016) | zebrafish | 2-mercaptobenzothiazole | TPO inhibition | 0-32 dpf | 20 dpf | 0.1, 0.35 mg/L | n/a | n/a | n/a | n/a | n/a | n/a | n/a | 0.35 mg/L | n/a | 3. 3.3, 7 Hig/L | | | 4 | 4 |
| Stinckens et al. (2016) | zebrafish | 2-mercaptobenzothiazole | TPO inhibition | 0-32 dpf | | 0.1, 0.35 mg/L | n/a | n/a | n/a | n/a | n/a | n/a | n/a | 0.35 mg/L | n/a | - | | | 1 | 1 |
| | zebrafish | 2-mercaptobenzothiazole | TPO inhibition | | 21 dpf | 0.1, 0.35 mg/L | | | n/a | | n/a | | n/a | | | - | | | 4 | 4 |
| | zebrafish | 2-mercaptobenzothiazole | TPO inhibition | 0-32 dpf 0-32 dpf | 22 dpf 23 dpf | 0.1, 0.35 mg/L | n/a n/a | n/a n/a | n/a n/a | n/a n/a | n/a n/a | n/a n/a | n/a n/a | 0.35 mg/L 0.35 mg/L | n/a n/a | - | | | £ | £ |
| | | | | | | | n/a | n/a | | | n/a | n/a | n/a | | n/a | - | | | 1 | .t |
| Stinckens et al. (2016) | zebrafish | 2-mercaptobenzothiazole | TPO inhibition | 0-32 dpf | 24 dpf | 0.1, 0.35 mg/L | | | n/a | n/a | | | | 0.35 mg/L | | - | | | - - | - |
| Stinckens et al. (2016) | zebrafish | 2-mercaptobenzothiazole | TPO inhibition | 0-32 dpf | 25 dpf | 0.1, 0.35 mg/L | n/a | n/a | n/a | n/a | n/a | n/a | n/a | 0.35 mg/L | n/a | - | | | | - |
| | zebrafish | 2-mercaptobenzothiazole | TPO inhibition | 0-32 dpf | 26 dpf | 0.1, 0.35 mg/L | n/a | n/a | n/a | n/a | n/a | n/a | n/a | 0.35 mg/L | 0.35 mg/L | - | | | | |
| Stinckens et al. (2016) | zebrafish | 2-mercaptobenzothiazole | TPO inhibition | 0-32 dpf | 27 dpf | 0.1, 0.35 mg/L | n/a | n/a | n/a | n/a | n/a | n/a | n/a | 0.35 mg/L | n/a | | | | | |
| Stinckens et al. (2016) | zebrafish | 2-mercaptobenzothiazole | TPO inhibition | 0-32 dpf | 28 dpf | 0.1, 0.35 mg/L | n/a | n/a | n/a | n/a | n/a | n/a | n/a | 0.35 mg/L | n/a | | | | | |
| Stinckens et al. (2016) | zebrafish | 2-mercaptobenzothiazole | TPO inhibition | 0-32 dpf | 29 dpf | 0.1, 0.35 mg/L | n/a | n/a | n/a | n/a | n/a | n/a | n/a | 0.35 mg/L | 0.35 mg/L | - | | | | |
| Stinckens et al. (2016) | zebrafish | 2-mercaptobenzothiazole | TPO inhibition | 0-32 dpf | 30 dpf | 0.1, 0.35 mg/L | n/a | n/a | n/a | n/a | n/a | n/a | n/a | 0.35 mg/L | 0.35 mg/L | - | | | - | - |
| Stinckens et al. (2016) | zebrafish | 2-mercaptobenzothiazole | TPO inhibition | 0-32 dpf | 31 dpf | 0.1, 0.35 mg/L | n/a | n/a | n/a | n/a | n/a | n/a | n/a | 0.35 mg/L | n/a | - | | | | |
| Stinckens et al. (2016) | zebrafish | 2-mercaptobenzothiazole | TPO inhibition | 0-32 dpf | 32 dpf | 0.1, 0.35 mg/L | n/a | n/a | n/a | n/a | 0.35 mg/L ² | - | n/a | 0.35 mg/L | n/a | | | | 2 | 2 |
| Nelson et al. (2016) | fathead minnow | 2-mercaptobenzothiazole | TPO inhibition | 0-21 dpf | 6 dpf | 0.25, 0.5, 1 mg/L | | n/a | n/a | n/a | 1 mg/L ^e | .t | - | n/a | n/a | - | | | | |
| Nelson et al. (2016) | fathead minnow | 2-mercaptobenzothiazole | TPO inhibition | 0-21 dpf | 14 dpf | 0.25, 0.5, 1 mg/L | 0.5, 1 mg/L* | n/a | n/a | 0.5, 1 mg/L ⁵ | n/a | 1 mg/L ^r | n/a | 0.5, 1 mg/L | n/a | - | | | n/a | ." |
| Nelson et al. (2016) | fathead minnow | 2-mercaptobenzothiazole | TPO inhibition | 0-21 dpf | 21 dpf | 0.25, 0.5, 1 mg/L | 1 mg/L* | n/a | n/a | 0.5, 1 mg/L ⁵ | .t | . ^t | n/a | 0.5, 1 mg/L | n/a | - | | | 0.25, 0.5, 1 mg/L ^c | ¹ . |
| Wei et al. (2018) | zebrafish | bisphenol S | unknown | adults | F1 96 hpf | 1, 10, 100 µg/L | n/a | n/a | n/a | n/a | 1, 10, 100 μg/L ^ε | | 1, 10, 100 μg/L | n/a | 1, 10, 100 µg/L | | | | | 1, 10, 100 μg/L ^ε |
| Crane et al. (2005) | fathead minnow | ammonium perchlorate | NIS inhibition | 0-28 dpf | 28 dpf | 1, 10, 100 mg/L | n/a | n/a | n/a | 1, 10, 100 mg/L ⁵ | _t | .ť | n/a | n/a | n/a | | | | 100 mg/L | |
| Crane et al. (2006) | fathead minnow | methimazole | TPO inhibition | 0-84 dpf | 28 dpf | 32, 100, 320 µg/L | n/a | n/a | n/a | n/a | 32, 100 μg/L ^ε | 320 μg/L ^ε | n/a | n/a | n/a | 32, 100 µg/L | | | <u>.</u> | . ^t |
| Crane et al. (2006) | fathead minnow | methimazole | TPO inhibition | 0-84 dpf | 56 dpf | 32, 100, 320 µg/L | n/a | n/a | n/a | n/a | .t | 100 μg/L ^ε | n/a | n/a | n/a | 32, 100 µg/L | | | 320 μg/L ^ε | .t |
| Crane et al. (2006) | fathead minnow | methimazole | TPO inhibition | 0-84 dpf | 84 dpf | 32, 100, 320 µg/L | n/a | n/a | n/a | n/a | | • | n/a | n/a | n/a | 32, 100 µg/L | | | | |
| Stinckens et al. (2020) | zebrafish | methimazole | TPO inhibition | 0-32 dpf | 21 dpf | 50, 100 mg/L | n/a | n/a | n/a | n/a | 50, 100 mg/L ² | 50, 100 mg/L [£] | | 50, 100 mg/L | n/a | | | | | |
| Stinckens et al. (2020) | zebrafish | methimazole | TPO inhibition | 0-32 dpf | 32 dpf | 50, 100 mg/L | n/a | n/a | n/a | n/a | 50, 100 mg/L ^c | 50, 100 mg/L ^c | | 50, 100 mg/L | 100 mg/L | | | | | |
| Stinckens et al. (2020) | zebrafish | propylthiouracil | TPO inhibition | 0-32 dpf | 14 dpf | 37, 111 mg/L | n/a | n/a | n/a | n/a | 37, 111 mg/L ^r | 111 mg/L ^r | | n/a | 111 mg/L | | | | | |
| Stinckens et al. (2020) | zebrafish | propylthiouracil | TPO inhibition | 0-32 dpf | 21 dpf | 37, 111 mg/L | n/a | n/a | n/a | n/a | 37, 111 mg/L ^E | 111 mg/L [£] | - | 37, 111 mg/L | 111 mg/L | | | | | |
| Stinckens et al. (2020) | zebrafish | propylthiouracil | TPO inhibition | 0-32 dpf | 32 dpf | 37, 111 mg/L | n/a | n/a | n/a | n/a | 37, 111 mg/L ^r | 37, 111 mg/L ^r | - | 37, 111 mg/L | - | | | | | |
| Stinckens et al. (2020) | zebrafish | iopanoic acid | DIO1 and 2 inhibition | 0-32 dpf | 9 dpf | 2 mg/L | n/a | n/a | n/a | n/a | n/a | n/a | 2 mg/L | n/a | n/a | 2 mg/L | | | | |
| Stinckens et al. (2020) | zebrafish | iopanoic acid | DIO1 and 2 inhibition | 0-32 dpf | 14 dpf | 0.35, 1 mg/L | n/a | n/a | n/a | n/a | .t | - ^t | - | n/a | 1, 2 mg/L | | | | | |
| Stinckens et al. (2020) | zebrafish | iopanoic acid | DIO1 and 2 inhibition | 0-32 dpf | 21 dpf | 0.35, 1 mg/L | n/a | n/a | n/a | n/a | _t | 0.35, 1 mg/L ^c | - | 0.35, 1, 2 mg/L | 0.35, 1, 2 mg/L | | | | | |
| Stinckens et al. (2020) | zebrafish | iopanoic acid | DIO1 and 2 inhibition | 0-32 dpf | 32 dpf | 0.35, 1, 2 mg/L 0, 50, 100, 150, 200, | n/a | n/a | n/a | n/a | 1. | 0.35, 1, 2 mg/L ^E | - | 0.35, 1, 2 mg/L | 0.35, 1, 2 mg/L | | | | | |
| Wang et al. (2020) | zebrafish | perfluorooctanoic acid (PFOA) | DIO1 and 2 inhibition | 0-5 dpf | 5 dpf | 2502, 300, 350, 400, 450, 500 mg/L 0, 400, 600, 800, 1000, 1200, 1400, 1600, 1800, | .• | - | 125, 250, 500 mg/L* | - | 250, 500 mg/L [£] | 250, 500 mg/L ^c | 200, 250, 300, 350, 400, 4 | i0 n/a | n/a | 300, 400, 450, 500 mg/L | | 500 mg/L | £ | <u>_</u> t |
| Wang et al. (2020) | zebrafish | PF030A | unknown | 0-5 dpf | 5 dpf | 2000, 2200, 2400 mg/L 0, 30, 45, 60, 90, 120, | 1200, 2200 mg/L* | .• | 600, 1200, 2200 mg/L* | - | 600, 1200, 2200 mg/L ^c | 1200, 2200 mg/L ^c | 800, 1000, 1200, 1400, 16 | | n/a | - | - | - | .t | . ⁴ |
| Wang et al. (2020) | zebrafish | PFO4DA | unknown | 0-5 dpf | 5 dpf | 150, 180, 210, 240 mg/L 0, 5, 10, 15, 20, 25, | .• | 240 mg/L* | .* | ÷ | | | α 45, 60, 90, 120, 150, 180, 3 | 1 n/a | n/a | - | - | - | . ¹ | . ¹ . |
| Wang et al. (2020) | zebrafish | PFO5DoDA | unknown | 0-5 dpf | 5 dpf | 30, 35, 40 mg/L | .* | | 10, 20, 40 mg/L* | | 10, 20, 40 mg/L ^c | 10, 20, 40 mg/L ^c | 20, 25, 30, 35, 40 mg/L ⁵ | n/a | n/a | | 10 mg/L | - | .t | . ^c |
| Rehberger et al. (2018) | | propylthiouracil | | 0-5 dpf | 5 dpf | 0, 2.5, 10, 25, 50 mg/L | | | n/a | 10, 25, 50 mg/L | n/a | | n/a | | n/a | n/a | | | | |

| Legend |
|---|
| n/a: not measured |
| * based on increased mRNA levels of the target as indirect measurement of MIE |
| \$ based on thyroid histopathology |
| £ based on whole body measurement |

§ based on visual evaluation of graphs because no statistics have been reported

References

Cavallin, J.E., Ankley, G.T., Blackwell, B.R., Blanksma, C.A., Fay, K.A., Jensen, K.M., Kahl, M.D., Knapen, D., Kosian, P.A., Poole, S.T., Randolph, E.C., Schroeder, A.L., Vergauwen, L., Villeneuve, D.L., 2017. Impaired swim bladder inflation in early life stage fathead minnows exposed to a deiodinase inhibitor, iopanoic acid. Environmental Toxicology and Chemistry 36, 2942-2952.

Crane, H.M., Pickford, D.B., Hutchinson, T.H., Brown, J.A., 2005. Effects of ammonium perchlorate on thyroid function in developing fathead minnows, Pimephales promelas. Environmental Health Perspectives 113, 396-401. Crane, H.M., Pickford, D.B., Hutchinson, T.H., Brown, J.A., 2006. The effects of methimazole on development of the fathead minnow, pimephales promelas, from embryo to adult. Toxicological Sciences 93, 278-285.

Nelson, K., Schroeder, A., Ankley, G., Blackwell, B., Blanksma, C., Degitz, S., Flynn, K., Jensen, K., Johnson, R., Kahl, M., Knapen, D., Kosian, P., Milsk, R., Randolph, E., Saari, T., Stinckens, E., Vergauwen, L., Villeneuve, D., 2016.

Impaired anterior swim bladder inflation following exposure to the thyroid peroxidase inhibitor 2-mercaptobenzothiazole part I: Fathead minnow. Aquatic Toxicology 173, 192-203. Rehberger, K., Baumann, L., Hecker, M., Braunbeck, T., 2018. Intrafollicular thyroid hormone staining in whole-mount zebrafish (Danio rerio) embryos for the detection of thyroid hormone synthesis disruption.

Fish Physiology and Biochemistry 44, 997-1010.

Stinckens, E., Vergauwen, L., Blackwell, B.R., Anldey, G.T., Villeneuve, D.L., Knapen, D., 2020. Effect of Thyroperoxidase and Deiodinase Inhibition on Anterior Swim Bladder Inflation in the Zebrafish. Environmental Science & Technology 54, 6213-6223.

Stinckens, E., Vergauwen, L., Schroeder, A., Maho, W., Blackwell, B., Witters, H., Blust, R., Ankley, G., Covaci, A., Villeneuve, D., Knapen, D., 2016.

Impaired anterior swim bladder inflation following exposure to the thyroid peroxidase inhibitor 2-mercaptobenzothiazole part II: Zebrafish. Aquatic Toxicology 173, 204-217.

Wang, J.X., Shi, G.H., Yao, J.Z., Sheng, N., Cui, R.N., Su, Z.B., Guo, Y., Dai, J.Y., 2020. Perfluoropolyether carboxylic acids (novel alternatives to PFOA) impair zebrafish posterior swim bladder development via thyroid hormone disruption. Environment International 134.

Wei, P.H., Zhao, F., Zhang, X.N., Liu, W.M., Jiang, G.B., Wang, H.F., Ru, S.G., 2018. Transgenerational thyroid endocrine disruption induced by bisphenol S affects the early development of zebrafish offspring. Environmental Pollution 243, 800-808.